

Thematic review series: *The Immune System and Atherogenesis*

Lipoprotein-associated inflammatory proteins: markers or mediators of cardiovascular disease?

Alan Chait,¹ Chang Yeop Han, John F. Oram, and Jay W. Heinecke

Division of Metabolism, Endocrinology, and Nutrition, Department of Medicine, University of Washington, Seattle, WA 98195

Abstract In humans, a chronically increased circulating level of C-reactive protein (CRP), a positive acute-phase reactant, is an independent risk factor for cardiovascular disease. This observation has led to considerable interest in the role of inflammatory proteins in atherosclerosis. In this review, after discussing CRP, we focus on the potential role in the pathogenesis of human vascular disease of inflammation-induced proteins that are carried by lipoproteins. Serum amyloid A (SAA) is transported predominantly on HDL, and levels of this protein increase markedly during acute and chronic inflammation in both animals and humans. Increased SAA levels predict the risk of cardiovascular disease in humans. Recent animal studies support the proposal that SAA plays a role in atherogenesis. Evidence is accruing that secretory phospholipase A₂, an HDL-associated protein, and platelet-activating factor acetylhydrolase, a protein associated predominantly with LDL in humans and HDL in mice, might also play roles both as markers and mediators of human atherosclerosis. In contrast to positive acute-phase proteins, which increase in abundance during inflammation, negative acute-phase proteins have received less attention. Apolipoprotein A-I (apoA-I), the major apolipoprotein of HDL, decreases during inflammation. Recent studies also indicate that HDL is oxidized by myeloperoxidase in patients with established atherosclerosis. These alterations may limit the ability of apoA-I to participate in reverse cholesterol transport. Para-oxonase-1 (PON1), another HDL-associated protein, also decreases during inflammation. PON1 is atheroprotective in animal models of hypercholesterolemia. Controversy over its utility as a marker of human atherosclerosis may reflect the fact that enzyme activity rather than blood level (or genotype) is the major determinant of cardiovascular risk. Thus, multiple lipoprotein-associated proteins that change in concentration during acute and chronic inflammation may serve as markers of cardiovascular disease. **In future studies, it will be important to determine whether these proteins play a causal role in atherogenesis.**—Chait, A., C. Y. Han, J. F. Oram, and J. W. Heinecke. **Lipoprotein-associated inflammatory proteins: markers or mediators of cardiovascular disease?** *J. Lipid Res.* 2005. 46: 389–403.

Supplementary key words inflammation • C-reactive protein • serum amyloid A • apolipoprotein A-I • high density lipoprotein • paraoxonase-1 • secretory phospholipase A₂ • platelet-activating factor acetylhydrolase • myeloperoxidase

In the face of infection, tissue damage, or acute inflammation, the host undergoes a series of biochemical and physiological changes termed the acute-phase response. This response plays a critical role in limiting tissue injury and in the innate immune response. The innate immune system provides a rapid, first-line defense against injurious insults, compared with the adaptive immune system, which uses slower and more specific B- and T-cell responses. The innate immune system uses a number of pattern recognition receptors that sense conserved structures and molecules characteristic of harmful agents. Pattern recognition receptors are important for triggering the activation of cells of the innate immune system and removing potentially injurious pathogens (1, 2).

A key component of the acute-phase response is altered hepatic synthesis of a wide array of proteins involved in coagulation, lipid metabolism, and the complement system (3). Various cytokines regulate changes in the concentrations of these proteins in blood. A low level of systemic inflammation can also chronically perturb blood levels of the inflammatory proteins that participate in the acute-phase response. Importantly, the magnitude of the chronic change is often considerably less than that observed in response to acute injury or infection. Moreover, many lines of evidence suggest that chronic, small changes in inflammatory protein levels might be detrimental rather than beneficial to the host.

The acute-phase reaction occurs rapidly in response to tissue injury or infection. For example, levels of C-reactive protein (CRP; so named because it reacts with the C-polysaccharide protein of *Streptococcus pneumoniae*) can increase

Manuscript received 16 December 2004 and in revised form 20 January 2005.

Published, JLR Papers in Press, February 1, 2005.
DOI 10.1194/jlr.R400017JLR200

¹ To whom correspondence should be addressed.
e-mail: achait@u.washington.edu

Copyright © 2005 by the American Society for Biochemistry and Molecular Biology, Inc.

This article is available online at <http://www.jlr.org>

1,000-fold within 48 h after an acute-phase stimulus. This is the largest increase achieved by any of the known acute-phase proteins. It is also important to note that concentrations of several proteins decrease in blood and the liver during the acute-phase response (3). These proteins are referred to as negative acute-phase proteins. A variety of functions have been attributed to the various positive and negative acute-phase proteins. For example, CRP activates the complement pathway and binds to Fc receptors on macrophages, which may be important for host defense.

Epidemiological and clinical studies have shown independent relationships between circulating levels of certain of these inflammatory proteins and cardiovascular disease, supporting the view that increased levels of CRP and other inflammatory proteins provide an independent assessment of the risk for cardiovascular events. These observations have reinforced basic science studies demonstrating that atherosclerosis is a chronic inflammatory process (2). Thus, one possibility is that altered levels of these inflammatory proteins reflect the presence of inflammation in the artery wall (4–6) or elsewhere in the body that might increase the risk for atherosclerosis. If so, these proteins might serve as potential markers of atherosclerosis. More recently, it has been proposed that inflammatory proteins may play a pathogenic role in atherosclerosis by promoting vascular injury. In this view, inflammatory proteins directly affect biological processes in the artery wall, or perhaps elsewhere, that in turn contribute to atherogenesis. This proposal raises the possibility that inflammatory proteins might be direct mediators of atherosclerosis.

In this review, we evaluate the roles of several positive and negative inflammatory proteins as markers or mediators of atherosclerosis. After discussing the role of CRP, we focus our attention on lipoprotein-associated proteins implicated in atherogenesis, including serum amyloid A (SAA), group II secretory phospholipase A₂ (sPLA₂), platelet-activating factor acetylhydrolase (PAF-AH), apolipoprotein J (apoJ; also known as clusterin), paraoxonase-1 (PON1), and apolipoprotein A-I (apoA-I). Several recent reviews provide information on the potential atherogenic role of non-lipoprotein-associated inflammatory proteins, such as complement, fibrinogen, ferritin, and ceruloplasmin (7–10).

ROLE OF PROTEINS INFLUENCED BY INFLAMMATION IN ATHEROGENESIS

CRP

CRP, a member of the pentraxin family of proteins, contains five noncovalently linked protomers surrounding a central pore. It is transported free in plasma rather than bound to circulating lipoproteins, although it can interact with oxidized phospholipids and oxidized lipoproteins in vitro (11). CRP activates complement and binds to Fc receptors, which may facilitate the uptake and clearance of apoptotic and necrotic cells during the acute-phase response (12).

Predictor of clinical cardiovascular events. CRP was the first acute-phase protein identified and thus is the best studied marker of inflammation in humans. Increased levels of CRP pre-

dict first clinical events, recurrent events, coronary heart disease end points, and stroke (reviewed in 13). In certain studies, CRP was a more powerful predictor of cardiovascular risk than traditional risk factors such as LDL (14). Indeed, CRP appeared to predict risk independently of LDL and HDL cholesterol (13, 15) and to provide predictive power beyond that derived from using Framingham risk scores (14). CRP levels are also increased in conditions that are associated with increased cardiovascular risk, including obesity (16–20), insulin resistance (17, 18), hypertension (21, 22), the metabolic syndrome (16, 23–25), type 2 diabetes (16, 17, 26), hypertriglyceridemia (24, 27, 28), a low level of HDL cholesterol (17, 24, 28), and smoking (28, 29). CRP levels are also increased in other chronic inflammatory conditions, such as periodontal disease (30, 31) and rheumatoid arthritis (32), which are more weakly associated with increased risk for cardiovascular disease. CRP is less well associated with measures of atherosclerosis than with clinical outcomes (33–35).

Several risk factors associated with increased CRP are also components of the metabolic syndrome, which greatly increases the risk of clinically significant atherosclerosis (36–39). Insulin resistance, diabetes, low HDL, visceral obesity, and hypertension are the hallmarks of the metabolic syndrome (40). Indeed, CRP adds prognostic information even when components of the metabolic syndrome are taken into account (41), although the magnitude of risk is attenuated after adjusting for these risk factors in some studies (42). However, the metabolic syndrome is unlikely to be the sole reason for the association between CRP and cardiovascular risk, because levels also are increased in conditions such as familial hypercholesterolemia (43), in which the prevalence of the metabolic syndrome is unlikely to be increased.

Although increased CRP has been regarded as one of the strongest predictors of risk for future cardiovascular events, the magnitude of this risk was recently called into question. In the Reykjavik Prospective Cohort Study (44), multivariate analysis suggested that measuring CRP added less to the predictive power than previous studies had suggested and less than traditional risk factors. After adjustment for factors such as smoking status, body mass index, and total blood cholesterol, participants in the highest tertile of CRP had only a 1.5-fold higher risk for coronary artery disease than those in the lowest tertile. In contrast, the odds ratio was 2.4 for subjects with increased cholesterol and 1.9 for smokers. Strengths of this study included the large number of subjects (>20,000, the largest number reported to date), the large number of cardiovascular events, the length of follow-up (20 years), and low rates of dropout in the study population (44). These observations suggest that CRP might contribute less to predictions than traditional risk factors of cardiovascular disease.

Two recent studies suggest that statins reduce CRP in patients with established cardiovascular disease and that clinical improvements in these patients is independent of changes in lipid levels (45, 46). These observations raise the possibility that inflammation plays a causal role in atherosclerosis and confirm that blood levels of CRP serve as

a marker for the risk of cardiovascular disease that is independent of traditional risk factors. However, the magnitude of this risk may have been overestimated in earlier studies, and uncertainties remain about the clinical utility of measuring CRP.

Atherogenic effects. Most of our understanding of the potential atherogenic mechanisms of CRP is based on model systems using cultured cells, especially endothelial cells. Proposed mechanisms include impaired production of cardioprotective molecules [e.g., endothelial nitric oxide (47) and prostacyclin (48)] and increased production of potentially atherogenic molecules [e.g., endothelin-1 (49), various cell adhesion molecules (47, 49, 50), monocyte chemoattractant protein 1 (51), interleukin-8 (52), and plasminogen activator inhibitor-1 (53)]. CRP induces macrophages to secrete tissue factor (54) and increases their production of reactive oxygen species (55) and proinflammatory cytokines (56). It also promotes monocyte adhesion and chemotaxis (57, 58), increases the uptake of oxidized LDL (11), and stimulates the expression of matrix metalloproteinases by macrophages (59). In vascular smooth muscle cells, CRP induces nitric oxide synthase, activates nuclear factor κ B and mitogen-activated protein kinase, and promotes cell proliferation (60). These latter effects may be partly attributable to upregulation of the angiotensin type-1 receptor (61). These studies have been reviewed recently (62).

It is important to note that these studies of cultured cells usually did not assess the purity of the CRP used, utilized high concentrations of the protein, and often failed to use relevant controls to demonstrate the specificity of the response. These are important issues because commercial CRP can be contaminated with other proteins and it can adopt a variety of properties in vitro that may not be physiologically relevant (63). However, if these in vitro studies are valid, CRP could be involved at multiple stages of early and late atherosclerosis: endothelial cell injury, impairment of vasodilation, enhanced monocyte adhesion and chemotaxis, lipid accumulation by monocyte-macrophages, smooth muscle cell proliferation, thrombosis, and plaque rupture. Moreover, CRP has been detected immunohistochemically in atherosclerotic lesions (11, 64, 65).

Remarkably little is known about CRP's effects in animal models of atherosclerosis. Because CRP is not an acute-phase protein in mice (66, 67), transgenic mouse models of accelerated atherosclerosis offer useful opportunities to directly test its atherogenic role. A recent study using apoE-deficient mice demonstrated that atherosclerosis was enhanced when animals overexpressed human CRP (68). However, the effects on lesion formation were modest and were confined to males, suggesting that even high levels of CRP might have little direct effect on the initiation and progression of atherosclerosis in hypercholesterolemic mice. In pioneering studies, Pepys and colleagues (69) demonstrated that human CRP promotes myocardial infarction in a rat model of acute ischemic injury. The increase in infarct size caused by CRP was abrogated by depleting complement in vivo, suggesting that complement activation by CRP plays a role in tissue injury.

CRP interacts with the complement and coagulation systems, and thrombosis is centrally important in triggering clinical events in human atherosclerotic vascular disease (70). Indeed, CRP colocalizes with activated complement in ischemic human myocardial tissue (71). In contrast, most animal models of hypercholesterolemia do not develop myocardial infarction or exhibit a major thrombotic component. The fact that CRP is more strongly associated with cardiovascular end points than with measures of atherosclerosis raises questions regarding whether its major effect is on thrombosis and plaque rupture rather than on atherogenesis.

Thus, although CRP might be an independent marker for cardiovascular disease risk, the role of CRP as a mediator of atherogenesis remains to be established. In future studies, it will be important to determine whether CRP directly promotes atherogenesis in animal models and to explore the possibility that it exacerbates vascular disease by other mechanisms, perhaps involving the complement and coagulation systems.

SAA

SAA is an amphipathic, α -helical apolipoprotein that is transported in the circulation primarily in association with HDL (72–74). Like CRP levels, SAA levels increase rapidly in the blood of humans suffering from acute inflammation (75). They also increase in response to inflammation in mice (75).

The SAA gene family clusters on chromosome 11 in humans and chromosome 7 in mice. In mice, there are four functional *Saa* genes (*Saa1* to *Saa4*). The liver produces SAA1 and SAA2 in response to systemic inflammation. Extrahepatic cells, such as macrophages and adipocytes, are the major source of SAA3, which also is produced in response to inflammatory stimuli (76, 77). SAA4 is produced in the liver constitutively (75). In humans, there are only three functional SAA genes, because *SAA3* is a pseudogene that is not transcribed. Although most SAA is produced by the liver, SAA1, SAA2, and SAA4 also can be produced by extrahepatic sources (78). It is of interest that mRNA for SAA has been detected in all of the major cell types present in atherosclerotic lesions (78), which contain both acute-phase and constitutive forms of SAA protein (79).

Predictor of clinical cardiovascular events. The concentration of SAA in blood increases dramatically during acute inflammation in humans and animals (75). In several observational and prospective studies, the risk of cardiovascular disease associated with SAA changed in parallel with that seen with CRP (80–83), although the absolute level of risk was generally smaller. Moreover, SAA levels are increased in conditions associated with increased cardiovascular risk, including obesity (17, 80), insulin resistance (17, 84), the metabolic syndrome (85), diabetes (17, 84, 86), and rheumatoid arthritis (32). Thus, a chronic modest increase in SAA level also appears to be associated with an increased risk of cardiovascular disease. Although additional studies are needed, these observations raise the possibility that SAA might serve as a marker for an increased risk of cardiovascular disease in humans.

Atherogenic effects. In vitro studies have suggested a number of pathways for the involvement of SAA in host defense mechanisms and inflammation. For example, in vitro experiments and studies in experimental animals indicate that apolipoprotein SAA can induce the expression of proteinases thought to degrade extracellular matrix (87, 88), which might be important during tissue injury. Moreover, it can act as a chemoattractant for inflammatory cells such as monocytes, polymorphonuclear leukocytes, and T-lymphocytes (89, 90), all of which are involved in host defense mechanisms. Lipoprotein-associated SAA may play a role in cholesterol transport by increasing the delivery of cholesterol to peripheral cells (91, 92), which might be important for lipid metabolism in injured cells. In contrast, liposome-associated SAA2 promotes the efflux of cholesterol from cells, a property not shared by SAA1 (93). Non-HDL-associated SAA promotes cholesterol efflux by both ABCA1-dependent and -independent mechanisms (94). When SAA circulates in the blood, it is bound to HDL, but it is conceivable that SAA not associated with lipoproteins could be formed from HDL in the artery wall or secreted directly by artery wall cells (78). Thus, lipoprotein-associated and non-lipoprotein-associated SAA might play different roles in the delivery and removal of cholesterol from cells at inflamed or injured sites. SAA has been shown to displace apoA-I from HDL in vitro (95). Moreover, remodeling of HDL occurs after the induction of acute inflammation with lipopolysaccharide (LPS) in mice (96). HDL protein composition also may be regulated by inflammation-induced changes in the hepatic expression of some HDL apolipoproteins, particularly SAA, apoA-I, and PON1 (see below). These compositional changes are likely to have functional consequences.

A rapid increase in SAA level is likely beneficial during infection and acute inflammation. However, modest but chronic increases of SAA, as often occur in the metabolic syndrome, type 2 diabetes, and other chronic inflammatory disorders, might be deleterious. Potential consequences include the stimulation of monocyte adhesion and chemotaxis into the artery wall and increased delivery of cholesterol to artery wall cells, two processes that might contribute to the initiation and progression of atherosclerotic lesions. Because SAA binds to proteoglycans (97, 98), chronic inflammation might facilitate the binding of SAA-containing HDL to extracellular vascular proteoglycans, which would favor the retention and modification of HDL by the vascular matrix. Retention of lipoproteins by vascular proteoglycans is believed to play an important role in the formation of macrophage foam cells but has also been implicated in all stages of atherogenesis (99, 100). Retention could prevent HDL from participating in reverse cholesterol transport and inhibiting oxidative processes in the artery wall. Moreover, modification of the lipid and protein components of trapped HDL might increase its interactions with macrophage scavenger receptors and render the lipoprotein atherogenic, similar to what happens with LDL (101).

Recent studies suggest that SAA may play a role in atherosclerosis in mouse models of hyperlipidemia. A study

of LDL receptor-deficient mice showed that supplementing a high-fat diet with cholesterol increases SAA levels without adversely affecting circulating lipid and lipoprotein concentrations (102). Circulating SAA levels in these mice, but not lipid levels, were strongly associated with the extent of atherosclerosis in the aorta. Binding of HDL to proteoglycans correlated with the SAA content of the lipoprotein, and SAA colocalized with apoA-I and proteoglycans in atherosclerotic lesions (102). Surprisingly, this study showed that a fraction of the SAA was carried in VLDL and its remnants in addition to HDL, the usual transporter of SAA through plasma (72, 74). These findings are consistent with the hypothesis that SAA might be atherogenic because it tethers SAA-containing lipoproteins to vascular proteoglycans. Its ability to stimulate the expression of matrix-degrading enzymes, such as collagenases and matrix metalloproteinases (88), also could contribute to plaque instability and plaque rupture, although this has not been tested in animal models.

These animal studies suggest that SAA is a mediator of atherosclerosis as well as a marker for cardiovascular disease. In future studies, it will be important to further explore the proposal that SAA plays a causal role in atherogenesis using additional animal models. One powerful test would be to overexpress the different SAA genes in hyperlipidemic mice that are not in an inflammatory state.

sPLA₂

sPLA₂ is another family of proteins that become more abundant in response to inflammatory stimuli. These enzymes hydrolyze phospholipids at the *sn*-2 position to generate lysophospholipids and free fatty acids. The best studied sPLA₂ with respect to its role in inflammation and atherogenesis is the group IIA enzyme. Like SAA and apoJ (see below), it also can be bound to HDL (103). Most studies of the role of sPLA₂ in atherogenesis have focused on the group IIA enzyme, and we will focus our review on this family member.

Predictor of clinical cardiovascular events. Circulating levels of sPLA₂ increase dramatically during infection and inflammation (104). Moreover, plasma levels are increased in patients with cardiovascular disease (105–108), consistent with a link between plasma sPLA₂ and atherosclerosis. However, it has yet to be determined whether this enzyme is an independent predictor of cardiovascular disease.

Atherogenic effects. In vitro studies indicate that sPLA₂ hydrolyzes phospholipids in both LDL and HDL. It converts LDL into small, dense particles of the type that are associated with an increased risk of cardiovascular disease in clinical studies (109, 110). Small, dense LDLs have an enhanced tendency to interact with proteoglycans (111). Moreover, sPLA₂ is also present in the artery wall, where it may act locally to promote the development of atherosclerosis. It is synthesized by vascular smooth muscle cells. Synthesis of the enzyme by vascular smooth muscle cells in vitro is induced by a variety of proinflammatory cytokines, although the specific cytokines involved appear to differ from smooth muscle cells of different species (105). In nonatherosclerotic arteries, sPLA₂ type IIA associates mainly

with smooth muscle cells (112, 113). In both humans and experimental animals, immunohistochemical studies have detected the enzyme in atherosclerotic lesions in association with macrophages, the necrotic lipid core, and extracellular vascular proteoglycans (112, 113), to which the enzyme binds with high affinity (114). The presence of sPLA₂ in the artery wall suggests that it might play a direct role in the development of vascular disease.

Several mechanisms might allow sPLA₂ to facilitate atherogenesis (reviewed in 105). When the enzyme modifies lipids of LDL and VLDL, two lipoproteins that contain apolipoprotein B-100, it generates lysophospholipids and free fatty acid. The resulting lipoproteins bind strongly to extracellular vascular proteoglycans (114). Once retained in the vascular matrix, these lipoproteins could undergo further enzymatic and nonenzymatic modifications, including oxidative modification. sPLA₂ also causes lipoproteins to aggregate and fuse (115), which further increases the binding of lipoproteins to extracellular proteoglycans (116). Aggregated lipoproteins can be taken up by LDL receptor-mediated phagocytosis (117–119). Because many molecules of lysophospholipids and free fatty acids can be generated when a single lipoprotein particle is hydrolyzed, high concentrations of these highly bioactive proinflammatory compounds might be achieved locally.

Lysophospholipids exert several potentially atherogenic effects. Lysophosphatidylcholine (lysoPC) is chemotactic for macrophages in vitro (120). Exposing vascular smooth muscle cells to lysoPC in vitro increased the expression of biglycan and the length of its glycosaminoglycan chains. These modified glycosaminoglycans increased the retention of unmodified LDL (121). Smooth muscle cells exposed to lysoPC also secreted the proteoglycan form of monocyte-macrophage colony-stimulating factor (M-CSF), the predominant proteoglycan produced by macrophages (121). M-CSF may play a role in lipoprotein retention and macrophage maturation and differentiation (122).

Endocytic uptake of lysoPC mediated by macrophage scavenger receptors plays a major role in oxidized LDL-induced macrophage growth (123, 124). LysoPC also can upregulate the expression of adhesion molecules for monocytes (125, 126) and stimulate the expression of growth factors, such as heparin binding epidermal growth factor and the platelet-derived growth factor A and B chains (127–129), all of which may play important roles in atherogenesis. Lysophosphatidic acid, which can be produced by hydrolysis of the polar head group of lysoPC, stimulates platelet aggregation, cell proliferation, and smooth muscle cell contraction (130, 131). Thus, phospholipid hydrolysis by sPLA₂ could produce high, localized concentrations of potentially toxic products in the milieu of the artery wall.

Mice in the C57Bl/6 genetic background, which are commonly used to study atherosclerosis, have a point mutation in the sPLA₂ gene that renders the enzyme nonfunctional (132). This defect ameliorates the reduction of HDL that normally occurs after an injection of LPS, a widely used model of acute inflammation. Transgenic mice that overexpress sPLA₂ are more susceptible to atherosclerosis than control mice (133, 134). Most likely, this partly

reflects the fact that overexpression of sPLA₂ reduces HDL levels in mice (135). Increased levels of sPLA₂ might similarly decrease HDL levels in humans suffering from acute or chronic inflammation. Transplanting bone marrow cells from sPLA₂ transgenic mice into LDL receptor-deficient mice also enhanced atherosclerosis (133), suggesting that the local presence of the enzyme in the artery wall milieu can facilitate atherogenesis, independently of its effect on HDL levels.

The role of sPLA₂ in mouse atherosclerosis has recently been questioned. The C57Bl/K mouse strain has an intact sPLA₂ gene (sPLA₂^{+/+}) and its genome is highly homologous to that of the C57Bl/6 mouse (136). When C57Bl/K mice were crossed with C57Bl/6 mice deficient in apoE (apoE^{-/-}), there were no differences in the extent of atherosclerosis (monitored by cholesteryl ester levels in the aorta) or HDL levels (quantified by fast-protein liquid chromatography) in siblings that were either apoE^{-/-} sPLA₂^{+/+} or apoE^{-/-} sPLA₂^{-/-} (136). Enzyme assays confirmed that there was no active sPLA₂ in the apoE^{-/-} sPLA₂^{-/-} mice. These observations raise questions about the impact of sPLA₂ on atherosclerosis in this animal model of hypercholesterolemia.

Several factors might account for the differences observed in the different mouse models of atherosclerosis. The transgenic mice that exhibited decreased HDL levels and increased atherosclerosis exhibited much higher blood levels of sPLA₂ than is observed in humans, raising questions about the physiological relevance of the model. The transgenic mouse studies also used the human enzyme, whereas the mouse enzyme was deficient in the apoE^{-/-} sPLA₂^{-/-} mice. It is noteworthy that mice in the C57Bl/6 genetic background, which have a point mutation in the sPLA₂ gene that renders the enzyme nonfunctional, are nonetheless susceptible to atherosclerosis. Finally, it is important to note that the C57Bl/K mice and the C57Bl/6 mice are not congenic strains and that other genetic differences also might contribute to their susceptibilities to atherosclerosis.

Collectively, these observations indicate that overexpression of the active enzyme in macrophages promotes atherosclerosis in hypercholesterolemic mice, raising the possibility that local expression of the enzyme in the artery wall might be atherogenic. It is less clear whether the enzyme plays a critical role in decreasing HDL levels during acute inflammation in mice and perhaps chronic inflammation in humans. Thus, these animal model studies suggest that sPLA₂ has the potential to promote vascular disease by a variety of mechanisms, raising the possibility that this enzyme directly mediates atherosclerosis in humans.

Secretory phospholipases other than group IIA sPLA₂ also may play a role in atherogenesis. Group V sPLA₂, which also can be increased by the injection of LPS, has proteoglycan-binding sites, facilitates LDL aggregation, and can promote foam cell formation in vitro (137). Moreover, it binds phosphatidylcholine more avidly than the group II enzymes and preferentially appears to hydrolyze phospholipids in HDL rather than LDL (138). By so doing, it may reduce the atheroprotective properties of HDL and render it potentially

atherogenic during inflammation. However, it is unclear whether this enzyme is normally present in atherosclerotic lesions and whether it plays a role in atherogenesis.

PAF-AH

PAF-AH (also known as lipoprotein-associated PLA₂) is another circulating PLA₂ that is transported in both LDL and HDL (139). In humans it is present mainly on LDL, whereas HDL is the preferred carrier in mice (reviewed in 140, 141). PAF-AH hydrolyzes both the acetyl group in the *sn*-2 position of PAF (a potent inflammatory mediator) and short-chain oxidized fatty acids in the *sn*-2 position of oxidized phospholipids. However, it is unclear whether PAF-AH is an acute-phase response protein, because studies have shown levels of the enzyme to be either increased or decreased after inflammatory stimuli in animal models (142, 143). Because of its ability to deactivate oxidized phospholipids in the potent proinflammatory and prothrombotic molecule PAF, the enzyme has been considered to have a potentially protective role in atherogenesis (144). However, epidemiological studies favor a proatherogenic role for the enzyme (see below), raising questions regarding its precise role in cardiovascular disease.

Predictor of clinical cardiovascular events. Despite the proposed antiatherogenic properties of PAF-AH, clinical studies have shown that an increased PAF-AH level is an independent predictor of cardiovascular risk (145–147). Moreover, PAF-AH levels do not correlate with CRP levels (145). Because of its uncertainty as a protein that changes during inflammation, it is unclear whether PAF-AH should be considered a marker for inflammatory cardiovascular disease, although high circulating levels appear to predict clinical events.

Atherogenic effects. PAF-AH has a potentially ambiguous role with respect to atherosclerosis. It might promote atherosclerosis by many of the same mechanisms proposed for sPLA₂. Conversely, it might be cardioprotective if it degrades oxidized phospholipids with atherogenic and thrombogenic properties. PAF, which is a substrate for PAF-AH, blocked the conversion of monocytes into migrating cells and favored their subendothelial retention in an in vitro model (148). Normal migration was restored by HDL-associated PAF-AH (149). It has been suggested that PAF-AH that is associated with HDL, as occurs predominantly in rodent species, might be an atheroprotective form of the enzyme, whereas LDL-associated PAF-AH, as occurs in humans, might be atherogenic (140).

Little is known about PAF-AH's influence on atherogenesis in animal models of vascular disease. However, reduced PAF-AH levels in apoE-deficient mice fed a Western diet were accompanied by an increase in circulating oxidized phospholipids (150). Transfer of the PAF-AH gene into skeletal muscle of apoE-deficient mice was associated with an increase in arterial wall thickness (151). Therefore, it remains to be firmly established whether the enzyme plays a causal role in cardiovascular disease.

PON1

The paraoxonases are a family of enzymes that protect cells from damage by organophosphate toxins (152, 153).

There are three known paraoxonase genes: *PON1*, *PON2*, and *PON3*. *PON1* is synthesized in the liver and transported through plasma in HDL (154). *PON2* is a ubiquitously expressed intracellular protein that can protect cells against oxidative damage (155). *PON3* is the least well studied. It also is transported in HDL (156, 157), but it has different substrate specificities than *PON1* (157). *PON1* activity and/or protein levels are inhibited during the acute-phase response in animals (158, 159) and also in humans (160). Other studies have demonstrated a reduction in paraoxonase expression in HepG2 cells exposed to oxidized LDL (161) and in livers of mice exposed to an atherogenic diet (162). In contrast, *PON3* does not appear to be regulated by either inflammation or exposure of liver cells to oxidized lipids (157).

Predictor of clinical cardiovascular events. Polymorphisms in *PON1* partly control the enzyme's activity in humans. An amino acid substitution at position 192 gives rise to two allozymes that have markedly different activities with certain artificial substrates in vitro. Some studies have found an association between the position 192 polymorphism and the risk for cardiovascular disease, but this association remains controversial (163–165).

It is noteworthy that *PON1* activity in serum varies widely by mechanisms that are poorly understood but are independent of the known polymorphisms. This has led to the proposal that enzyme activity rather than genotype plays a role in cardiovascular disease. Case-control and prospective studies of humans support this hypothesis. Thus, *PON1* may also serve as a marker for cardiovascular disease risk in humans, but enzyme activity rather than genotype or protein level appears to correlate most highly with the degree of risk (164).

Atheroprotective effects. In vitro studies suggest that HDL-associated *PON1* inhibits lipid peroxidation or degrades biologically active oxidized lipids in LDL (143, 166–170). LDL oxidation is thought to be one important mechanism for converting the lipoprotein to a form that promotes the formation of lipid-laden macrophages, the cellular hallmark of the early atherosclerotic lesion (171). HDL inhibits LDL oxidation by metal ions in vitro (172), leading to the suggestion that this pathway represents one potential cardioprotective function of HDL.

A deficiency of *PON1* enhanced atherosclerosis in hypercholesterolemic mice and was associated with an increase in oxidized phospholipids (173). Cell culture studies suggest that LDL isolated from these animals is enriched in potentially atherogenic oxidized lipids (174). Moreover, mice that overexpress *PON1* appear to be protected from atherosclerosis, and LDL isolated from these animals appears to contain less oxidized lipid (175). Collectively, these observations led to the suggestion that HDL-associated *PON1* prevents atherosclerosis by inhibiting lipid oxidation (166, 176). They also strongly support the proposal that HDL can affect atherosclerosis by a mechanism independent of reverse cholesterol transport.

Although *PON1* clearly has atheroprotective properties in animal models of hypercholesterolemia (173, 175), recent studies have raised questions regarding the precise mechanism of this effect. For example, low levels of PAF-AH were

detected in purified PON1, and biochemical studies suggested that PAF-AH was responsible for degrading biologically active oxidized lipids (177). Thus, the phospholipase activity of PON1 is not likely to be the main basis for the atheroprotection observed in the genetic mouse models. Moreover, the role of PON1 in the inhibition of LDL oxidation catalyzed by copper or peroxy radical has been questioned (178). Thus, the precise physiological function of PON1 remains unclear. Nonetheless, the genetic experiments in mice clearly indicate that it has atheroprotective functions.

ApoJ

ApoJ, also known as clusterin, travels through plasma while bound to HDL, and its level increases in response to inflammatory stimuli (179). Although little is known about its precise function, its presence in atherosclerotic lesions but not in normal arteries (180) suggests a role in the atherosclerosis associated with chronic inflammatory states. Its colocalization in lesions with apoA-I and apoE (180) suggests that it may enter lesions in association with HDL particles, which can be retained by vascular proteoglycans if they contain positively charged apolipoproteins such as apoE (181, 182) or SAA (102). Like SAA, apoJ can be synthesized by vascular cells, especially arterial smooth muscle cells and foam cells (180). It also is released during the platelet aggregation (183) that occurs during thrombosis and plaque rupture.

Predictor of clinical cardiovascular events. Circulating levels of apoJ increase with aging, in inflammatory states, and in diabetes (184, 185), suggesting that this apolipoprotein may be a potential marker for atherosclerosis in humans. Two studies, however, have failed to find a significant association between plasma apoJ levels and risk for coronary events (186, 187). Thus, it appears unlikely that apoJ is a strong independent predictor of cardiovascular disease.

Atherogenic effects. In animals, apoJ levels increase in situations in which PON1 levels decrease. For example, the apoJ/PON1 ratio is increased by feeding mice an atherogenic diet, by injecting mildly oxidized LDL into mice that are susceptible to atherosclerosis, or by inducing inflammation in rabbits (161). Exposing hepatocytes to mildly oxidized LDL also promotes apoJ expression and PON1 suppression in vitro (161). In a small clinical study, the apoJ/PON1 ratio in patients with coronary artery disease predicted whether HDL would protect LDL from becoming oxidized and inducing monocyte adherence and chemotaxis in vitro (161). These results suggest that apoJ may have a deleterious effect on the antioxidation activity of PON1, although the latter has recently been called into question (178).

These human and animal model studies have not yet established a role for apoJ in either facilitating or protecting against human atherosclerosis. Studies using mice with altered apoJ expression would provide a powerful tool for determining the influence of this protein on atherogenesis.

ApoA-I

Although apoA-I, HDL's major protein, is not widely regarded as an inflammatory molecule, apoA-I levels clearly

decrease during the acute inflammatory response in rabbits (188), mice (135), and humans (189). HDL is one of the major blood components that bind to bacterial LPS (190), and high HDL levels protect animals from LPS-induced septic shock (191). Studies suggest that HDL also has potent anti-inflammatory and antioxidant properties (reviewed in 166, 170), although the mechanisms are incompletely understood.

Predictor of clinical cardiovascular events. Numerous population studies have demonstrated an inverse relationship between plasma HDL levels and cardiovascular disease risk, establishing low HDL levels as a strong independent marker for atherosclerosis (192–194). Both the hepatic production rate and peripheral catabolism of apoA-I can influence plasma levels of HDL, and it is unclear to what extent these two different mechanisms contribute to low HDL levels during inflammation. It is noteworthy, however, that a low HDL level is one of the features of the metabolic syndrome (195), a condition strongly associated with increased levels of CRP and an increased risk of cardiovascular disease. The acute-phase reactant, SAA, can displace apoA-I from HDL in vitro (95). Moreover, HDL is remodeled to a considerable extent during inflammation (96). Such compositional changes are likely to be associated with altered functional properties. Therefore, it is likely that many atherogenic inflammatory states are associated with low plasma HDL and apoA-I levels and with altered HDL composition.

In addition to its plasma concentrations, apoA-I may be a marker for atherosclerosis when it is oxidized. Chlorinated HDL and nitrated HDL appear in human atherosclerotic lesions (196–199), and in vitro experiments indicate that these abnormal forms of the lipoprotein arise when apoA-I is oxidized by myeloperoxidase (200), a heme enzyme produced by macrophages. Moreover, tandem mass spectrometric analysis identified myeloperoxidase as a component of lesion HDL (196), suggesting that the enzyme and lipoprotein interact in the artery wall. Protein chlorination and nitration were impaired in myeloperoxidase-deficient mice (201, 202), strongly implicating myeloperoxidase in generating chlorinating and nitrating intermediates in vivo. 3-Chlorotyrosine and 3-nitrotyrosine were also detected in circulating HDL, and the levels of these oxidized amino acids were increased in HDL isolated from the blood of humans with established coronary artery disease (196–198). These observations raise the possibility that circulating levels of chlorinated and nitrated HDL are novel markers for clinically significant atherosclerosis.

Atheroprotective effects. Many lines of evidence indicate that HDL protects the artery wall from atherosclerosis. In one important pathway, apoA-I removes cellular cholesterol and phospholipids by an active transport process mediated by ABCA1 (203–205). In vivo, ABCA1 is critical for removing cholesterol from macrophages, and lipid-laden macrophages are the cellular hallmark of atherosclerotic lesions. HDL particles are also capable of clearing excess cholesterol from macrophages through their interaction with other cell surface proteins, such as scavenger recep-

tor B1 and ABCG1. The cardioprotective effects of HDL may occur by other mechanisms, including inhibiting LDL oxidation (172), reducing LDL lipid hydroperoxides (143) and oxidized phospholipids (170), and transporting oxidized lipids to the liver for elimination in the bile (206). HDL can also block the expression of adhesion molecules on endothelial cells (207, 208), thus blocking the recruitment of monocytes into the artery wall. HDL also can neutralize CRP's proinflammatory activity (209). It is likely that both lipids and proteins in HDL mediate these diverse atheroprotective effects.

The atheroprotective effects of HDL may be uniquely targeted for damage by inflammatory pathways. As discussed above, remodeling of HDL by SAA could convert it into an atherogenic particle. HDL or apoA-I exposed to HOCl or myeloperoxidase (the only enzyme known to generate HOCl in humans) loses its ability to remove cholesterol from cultured cells by the ABCA1 pathway (196). Impaired ABCA1 activity strongly associates with apoA-I chlorination but not nitration (196, 197, 210). It is noteworthy that bacterial products and cytokines are potent stimulants for oxidant generation by macrophages and other phagocytic white blood cells. It is possible, therefore, that HDL is targeted for oxidation in chronic inflammation and that HDL chlorination promotes atherosclerosis by inhibiting ABCA1-dependent cholesterol efflux from cells of the artery wall.

Taken together, these studies indicate that HDL could have direct anti-inflammatory effects that protect the artery wall from atherogenesis and that multiple inflammatory processes that alter HDL or apoA-I could play a direct role in atherogenesis, either by generating atherogenic particles or by impairing cholesterol efflux from arterial macrophages. It will be important to confirm these observations in larger clinical studies and with additional animal models.

REGULATION OF HEPATIC PRODUCTION OF PROTEINS DURING INFLAMMATION

Levels of CRP and SAA tend to increase in parallel during the acute inflammatory response, in chronic inflammatory conditions, and in conditions associated with an increased risk of cardiovascular disease (13, 75, 80–83). Therefore, it is not surprising that their synthesis and secretion by the liver are regulated in a similar manner. Both CRP and the inflammatory forms of SAA are secreted by the liver primarily in response to inflammatory stimuli such as interleukin-6, tumor necrosis factor- α , and interleukin-1 (12, 75). All of these cytokines are produced by macrophages (reviewed in 75). However, other cell types, particularly adipocytes (211, 212), can also synthesize and release these proteins. Indeed, it has been suggested that cytokines derived from adipose tissue are the major source of the increased levels of inflammatory markers seen in obesity (29, 213, 214). The recent suggestion that macrophages accumulate in adipose tissue of obese mice and humans (211, 212) may change our understanding of the role of these cells in generating cytokines and other in-

flammatory mediators in adipose tissue. The 3T3 adipocyte cell line also can secrete SAA directly (215). Moreover, macrophages present in adipose tissue from obese subjects may be a direct source of cytokines that induce the hepatic expression of molecules such as CRP and SAA or may secrete proteins such as SAA directly (75). These observations suggest an additional potential mechanism linking obesity, inflammation, and atherosclerosis.

Many other inflammatory stimuli prompt macrophages and other cells to generate cytokines, which in turn could result in the overproduction in the liver of acute-phase proteins such as CRP and SAA. One potential mechanism involves chronic infection with bacterial or viral pathogens, including *Chlamydia pneumoniae* (216–218), herpes simplex virus (217, 219), cytomegalovirus (217, 218), and *Helicobacter pylori* (218). However, epidemiological studies and intervention studies with antibiotics have yielded conflicting results regarding the role of chronic infection in atherosclerosis. However, cytokine levels are also increased in chronic inflammatory conditions such as periodontal disease, rheumatoid arthritis, and systemic lupus erythematosus. Under these conditions, cytokines might also induce increases of inflammatory proteins that promote atherosclerotic vascular disease.

CRP and SAA levels are increased in patients with the metabolic syndrome (16, 17, 24, 41). Their production in the liver could be stimulated by cytokines from the increased visceral adipose tissue that characterizes this disorder. As noted above, macrophages in adipose tissue may be a source of the cytokines that stimulate the hepatic production of inflammatory proteins or may even secrete some of these proteins directly. Moreover, adipocytes themselves can be the source of cytokines (211) and inflammatory molecules (215). Insulin resistance has also been proposed to regulate the levels of inflammatory molecules, although the mechanism is unclear. However, it is difficult to separate the role of insulin resistance from that of obesity, because the two are clearly interdependent. Nonetheless, subjects whose insulin resistance decreased when they lost weight had decreased levels of circulating CRP and SAA, whereas there was little change in these acute-phase proteins in subjects who lost weight but did not increase their insulin sensitivity (18). These observations raise the question of whether insulin resistance may regulate the circulating levels of inflammatory molecules independently of their association with obesity.

A wide variety of enzymes involved in hepatic lipid metabolism are regulated during acute inflammation. Indeed, a decreased level of blood cholesterol, together with variable changes in triglyceride levels, is one of the metabolic hallmarks of acute inflammation (220). Array analyses of mRNAs isolated from mice injected with LPS demonstrated coordinated downregulation of enzymes involved in cholesterol synthesis, triglyceride synthesis, and mitochondrial fatty acid oxidation (221). These observations indicate that acute inflammation affects levels of proatherogenic and antiatherogenic lipoproteins, raising the possibility that chronic inflammation also alters levels of these lipoproteins.

In humans, apoA-I and HDL levels are also reduced during acute inflammation (189) as well as in conditions associated with chronic inflammation, such as obesity and the metabolic syndrome (222–224). HDL levels are also low in patients suffering from type 2 diabetes, a major risk factor for accelerated vascular disease. LPS and the inflammatory cytokines interleukin-1, tumor necrosis factor- α , and interleukin-6 inhibit apoA-I expression by the liver (225, 226). Moreover, these cytokines resulted in reciprocal regulation of the expression of apoA-I and SAA in murine hepatocytes, such that apoA-I expression was inhibited and SAA expression was coordinately increased (226). These effects appear to be regulated by nuclear factor κ B, which inhibits the peroxisome proliferator-activated receptor α (225, 226). The net effect of inflammation may be to reduce HDL's ability to participate in reverse cholesterol transport and to perform its other atheroprotective functions.

CONCLUSIONS

There is mounting evidence that the chronic increase of proteins that participate in the acute inflammatory response may not only predict the onset of clinical cardiovascular events but may actually play a causal role in mediating atherosclerotic disease. The best studied of these is CRP. Increased levels of CRP appear to be an independent risk factor for cardiovascular disease, although the absolute magnitude of this risk remains to be established. Basic studies have identified a number of potential roles for CRP in atherogenesis, but additional studies need to be undertaken to determine whether CRP directly promotes vascular disease. SAA levels also appear to predict the risk of cardiovascular disease in humans, and recent animal studies support the proposal that this acute-phase protein plays a role in atherogenesis. It will be important to further investigate its potential role as a marker or mediator of atherosclerotic vascular disease. Less is known about the roles of sPLA₂, PAF-AH, and apoJ in atherogenesis, although evidence is accruing that sPLA₂ and PAF-AH also might play roles in the pathogenesis of atherosclerosis.

Negative acute-phase proteins such as apoA-I and PON1 have received less attention as markers of inflammation than have positive acute-phase proteins such as CRP and SAA. ApoA-I, the major apolipoprotein of HDL, decreases during both acute and chronic inflammation. It is well established that low levels of HDL are a major risk factor for accelerated atherosclerosis. Recent studies strongly suggest that levels of chlorinated and nitrated HDL are increased in patients with established cardiovascular disease. In vitro studies suggest that the ability of chlorinated apoA-I to promote cholesterol efflux by the ABCA1 pathway is markedly impaired. In larger clinical studies, it will be of great interest to examine the potential of oxidized HDL to serve as a marker for cardiovascular disease. PON1, another HDL-associated protein, also decreases during inflammation. It is clearly atheroprotective in animal models of hypercholesterolemia, and controversy over its util-

ity as a marker of human atherosclerosis may reflect the fact that enzyme activity rather than blood level (or genotype) is the major determinant of cardiovascular risk. The underlying mechanism may involve the degradation of oxidized phospholipids by PON1. However, recent studies have raised questions regarding this proposal. It will be important to clarify the underlying mechanism whereby PON1 protects animals from atherosclerosis.

Thus, many proteins that participate in the acute-phase response are implicated in the diagnosis and pathogenesis of clinically significant atherosclerotic vascular disease. By better understanding the pathophysiology of these proteins, we will begin to identify the pathways that promote atherosclerosis. These findings should provide new clinical tools for identifying patients at risk for cardiovascular disease and identify therapies that act by mechanisms that differ from those of traditional lipid-lowering therapies. **■**

This work was supported in part by National Institutes of Health Grants HL-30086, HL-018645, DK-02456, ES-07033, HL-075340, AG-021191, and by a grant from the Donald W. Reynolds Foundation.

REFERENCES

1. Janeway, C. A., Jr., and R. Medzhitov. 2002. Innate immune recognition. *Annu. Rev. Immunol.* **20**: 197–216.
2. Hansson, G. K., P. Libby, U. Schonbeck, and Z. Q. Yan. 2002. Innate and adaptive immunity in the pathogenesis of atherosclerosis. *Circ. Res.* **91**: 281–291.
3. Gabay, C., and I. Kushner. 1999. Acute-phase proteins and other systemic responses to inflammation. *N. Engl. J. Med.* **340**: 448–454.
4. Ross, R. 1993. The pathogenesis of atherosclerosis: a perspective for the 1990s. *Nature.* **362**: 801–809.
5. Libby, P. 2002. Inflammation in atherosclerosis. *Nature.* **420**: 868–874.
6. Ross, R. 1999. Atherosclerosis—an inflammatory disease. *N. Engl. J. Med.* **340**: 115–126.
7. Oksjoki, R., P. T. Kovanen, and M. O. Pentikainen. 2003. Role of complement activation in atherosclerosis. *Curr. Opin. Lipidol.* **14**: 477–482.
8. Reinhart, W. H. 2003. Fibrinogen—marker or mediator of vascular disease? *Vasc. Med.* **8**: 211–216.
9. de Valk, B., and J. J. Marx. 1999. Iron, atherosclerosis, and ischemic heart disease. *Arch. Intern. Med.* **159**: 1542–1548.
10. Fox, P. L., B. Mazumder, E. Ehrenwald, and C. K. Mukhopadhyay. 2000. Ceruloplasmin and cardiovascular disease. *Free Radic. Biol. Med.* **28**: 1735–1744.
11. Chang, M. K., C. J. Binder, M. Torzewski, and J. L. Witztum. 2002. C-reactive protein binds to both oxidized LDL and apoptotic cells through recognition of a common ligand: phosphorylcholine of oxidized phospholipids. *Proc. Natl. Acad. Sci. USA.* **99**: 13043–13048.
12. Volanakis, J. E. 2001. Human C-reactive protein: expression, structure, and function. *Mol. Immunol.* **38**: 189–197.
13. Pearson, T. A., G. A. Mensah, R. W. Alexander, J. L. Anderson, R. O. Cannon 3rd, M. Criqui, Y. Y. Fadl, S. P. Fortmann, Y. Hong, G. L. Myers, N. Rifai, S. C. Smith, Jr., K. Taubert, R. P. Tracy, and F. Vinicor. 2003. Markers of inflammation and cardiovascular disease: application to clinical and public health practice. A statement for healthcare professionals from the Centers for Disease Control and Prevention and the American Heart Association. *Circulation.* **107**: 499–511.
14. Ridker, P. M., N. Rifai, L. Rose, J. E. Buring, and N. R. Cook. 2002. Comparison of C-reactive protein and low-density lipoprotein cholesterol levels in the prediction of first cardiovascular events. *N. Engl. J. Med.* **347**: 1557–1565.
15. Albert, M. A., R. J. Glynn, and P. M. Ridker. 2003. Plasma concen-

- tration of C-reactive protein and the calculated Framingham Coronary Heart Disease Risk Score. *Circulation*. **108**: 161–165.
16. Guerrero-Romero, F., and M. Rodriguez-Moran. 2003. Relation of C-reactive protein to features of the metabolic syndrome in normal glucose tolerant, impaired glucose tolerant, and newly diagnosed type 2 diabetic subjects. *Diabetes Metab.* **29**: 65–71.
 17. Leinonen, E., E. Hurt-Camejo, O. Wiklund, L. M. Hulten, A. Hukka, and M. R. Taskinen. 2003. Insulin resistance and adiposity correlate with acute-phase reaction and soluble cell adhesion molecules in type 2 diabetes. *Atherosclerosis*. **166**: 387–394.
 18. McLaughlin, T., F. Abbasi, C. Lamendola, L. Liang, G. Reaven, P. Schaaf, and P. Reaven. 2002. Differentiation between obesity and insulin resistance in the association with C-reactive protein. *Circulation*. **106**: 2908–2912.
 19. Visser, M., L. M. Bouter, G. M. McQuillan, M. H. Wener, and T. B. Harris. 1999. Elevated C-reactive protein levels in overweight and obese adults. *J. Am. Med. Assoc.* **282**: 2131–2135.
 20. Weiss, R., J. Dziura, T. S. Burgert, W. V. Tamborlane, S. E. Taksali, C. W. Yeckel, K. Allen, M. Lopes, M. Savoye, J. Morrison, R. S. Sherwin, and S. Caprio. 2004. Obesity and the Metabolic Syndrome in Children and Adolescents. *Obstet. Gynecol. Surv.* **59**: 822–824.
 21. Niskanen, L., D. E. Laaksonen, K. Nyyssonen, K. Punnonen, V. P. Valkonen, R. Fuentes, T. P. Tuomainen, R. Salonen, and J. T. Salonen. 2004. Inflammation, abdominal obesity, and smoking as predictors of hypertension. *Hypertension*. **44**: 859–865.
 22. Bautista, L. E., L. M. Vera, I. A. Arenas, and G. Gamarra. 2005. Independent association between inflammatory markers (C-reactive protein, interleukin-6, and TNF-alpha) and essential hypertension. *J. Hum. Hypertens.* **19**: 149–154.
 23. Festa, A., R. D'Agostino, Jr., G. Howard, L. Mykkanen, R. P. Tracy, and S. M. Haffner. 2000. Chronic subclinical inflammation as part of the insulin resistance syndrome: the Insulin Resistance Atherosclerosis Study (IRAS). *Circulation*. **102**: 42–47.
 24. Tamakoshi, K., H. Yatsuya, T. Kondo, Y. Hori, M. Ishikawa, H. Zhang, C. Murata, R. Otsuka, S. Zhu, and H. Toyoshima. 2003. The metabolic syndrome is associated with elevated circulating C-reactive protein in healthy reference range, a systemic low-grade inflammatory state. *Int. J. Obes. Relat. Metab. Disord.* **27**: 443–449.
 25. Aronson, D., R. Sella, M. Sheikh-Ahmad, A. Kerner, O. Avizohar, S. Rispler, P. Bartha, W. Markiewicz, Y. Levy, and G. J. Brook. 2004. The association between cardiorespiratory fitness and C-reactive protein in subjects with the metabolic syndrome. *J. Am. Coll. Cardiol.* **44**: 2003–2007.
 26. Mojiminiyi, O. A., N. Abdella, M. A. Moussa, A. O. Akanji, H. Al Mohammedi, and M. Zaki. 2002. Association of C-reactive protein with coronary heart disease risk factors in patients with type 2 diabetes mellitus. *Diabetes Res. Clin. Pract.* **58**: 37–44.
 27. Grau, A. J., F. Bugge, H. Becher, E. Werle, and W. Hacke. 1996. The association of leukocyte count, fibrinogen and C-reactive protein with vascular risk factors and ischemic vascular diseases. *Thromb. Res.* **82**: 245–255.
 28. Fredrikson, G. N., B. Hedblad, J. A. Nilsson, R. Alm, G. Berglund, and J. Nilsson. 2004. Association between diet, lifestyle, metabolic cardiovascular risk factors, and plasma C-reactive protein levels. *Metabolism*. **53**: 1436–1442.
 29. McCarty, M. F. 1999. Interleukin-6 as a central mediator of cardiovascular risk associated with chronic inflammation, smoking, diabetes, and visceral obesity: down-regulation with essential fatty acids, ethanol and pentoxifylline. *Med. Hypotheses*. **52**: 465–477.
 30. Slade, G. D., E. M. Ghezzi, G. Heiss, J. D. Beck, E. Riche, and S. Offenbacher. 2003. Relationship between periodontal disease and C-reactive protein among adults in the Atherosclerosis Risk in Communities Study. *Arch. Intern. Med.* **163**: 1172–1179.
 31. Joshipura, K. J., H. C. Wand, A. T. Merchant, and E. B. Rimm. 2004. Periodontal disease and biomarkers related to cardiovascular disease. *J. Dent. Res.* **83**: 151–155.
 32. Wong, M., L. Toh, A. Wilson, K. Rowley, C. Karschimkus, D. Prior, E. Romas, L. Clemens, G. Dragicevic, H. Harianto, I. Wicks, G. McColl, J. Best, and A. Jenkins. 2003. Reduced arterial elasticity in rheumatoid arthritis and the relationship to vascular disease risk factors and inflammation. *Arthritis Rheum.* **48**: 81–89.
 33. Folsom, A. R., J. S. Pankow, R. P. Tracy, D. K. Arnett, J. M. Peacock, Y. Hong, L. Djousse, and J. H. Eckfeldt. 2001. Association of C-reactive protein with markers of prevalent atherosclerotic disease. *Am. J. Cardiol.* **88**: 112–117.
 34. Redberg, R. F., N. Rifai, L. Gee, and P. M. Ridker. 2000. Lack of association of C-reactive protein and coronary calcium by electron beam computed tomography in postmenopausal women: implications for coronary artery disease screening. *J. Am. Coll. Cardiol.* **36**: 39–43.
 35. Hunt, M. E., P. G. O'Malley, M. N. Vernalis, I. M. Feuerstein, and A. J. Taylor. 2001. C-reactive protein is not associated with the presence or extent of calcified subclinical atherosclerosis. *Am. Heart J.* **141**: 206–210.
 36. Reaven, G. M. 1994. Syndrome X: 6 years later. *J. Intern. Med. Suppl.* **736**: 13–22.
 37. Lamarche, B., I. Lemieux, and J. P. Despres. 1999. The small, dense LDL phenotype and the risk of coronary heart disease: epidemiology, patho-physiology and therapeutic aspects. *Diabetes Metab.* **25**: 199–211.
 38. Despres, J. P. 1993. Abdominal obesity as important component of insulin-resistance syndrome. *Nutrition*. **9**: 452–459.
 39. Despres, J.-P., S. Moorjani, A. Tremblay, M. Ferland, P. J. Lupien, A. Nadeau, and C. Bouchard. 1989. Relation of high plasma triglyceride levels associated with obesity and regional adipose tissue distribution to plasma lipoprotein-lipid composition in premenopausal women. *Clin. Invest. Med.* **12**: 374–380.
 40. Reaven, G. M. 1993. Role of insulin resistance in human disease (syndrome X): an expanded definition. *Annu. Rev. Med.* **44**: 121–131.
 41. Ridker, P. M., J. E. Buring, N. R. Cook, and N. Rifai. 2003. C-reactive protein, the metabolic syndrome, and risk of incident cardiovascular events: an 8-year follow-up of 14 719 initially healthy American women. *Circulation*. **107**: 391–397.
 42. Pirro, M., J. Bergeron, G. R. Dagenais, P. M. Bernard, B. Cantin, J. P. Despres, and B. Lamarche. 2001. Age and duration of follow-up as modulators of the risk for ischemic heart disease associated with high plasma C-reactive protein levels in men. *Arch. Intern. Med.* **161**: 2474–2480.
 43. van Wissen, S., M. D. Trip, T. J. Smilde, J. de Graaf, A. F. Stalenhoef, and J. J. Kastelein. 2002. Differential hs-CRP reduction in patients with familial hypercholesterolemia treated with aggressive or conventional statin therapy. *Atherosclerosis*. **165**: 361–366.
 44. Danesh, J., J. G. Wheeler, G. M. Hirschfield, S. Eda, G. Eiriksdottir, A. Rumley, G. D. Lowe, M. B. Pepys, and V. Gudnason. 2004. C-reactive protein and other circulating markers of inflammation in the prediction of coronary heart disease. *N. Engl. J. Med.* **350**: 1387–1397.
 45. Ridker, P. M., C. P. Cannon, D. Morrow, N. Rifai, L. M. Rose, C. H. McCabe, M. A. Pfeffer, and E. Braunwald. 2005. C-reactive protein levels and outcomes after statin therapy. *N. Engl. J. Med.* **352**: 20–28.
 46. Nissen, S. E., E. M. Tuzcu, P. Schoenhagen, T. Crowe, W. J. Sasiela, J. Tsai, J. Orazem, R. D. Magorien, C. O'Shaughnessy, and P. Ganz. 2005. Statin therapy, LDL cholesterol, C-reactive protein, and coronary artery disease. *N. Engl. J. Med.* **352**: 29–38.
 47. Venugopal, S. K., S. Devaraj, I. Yuhanna, P. Shaul, and I. Jialal. 2002. Demonstration that C-reactive protein decreases eNOS expression and bioactivity in human aortic endothelial cells. *Circulation*. **106**: 1439–1441.
 48. Venugopal, S. K., S. Devaraj, and I. Jialal. 2003. C-reactive protein decreases prostacyclin release from human aortic endothelial cells. *Circulation*. **108**: 1676–1678.
 49. Verma, S., S. H. Li, M. V. Badiwala, R. D. Weisel, P. W. Fedak, R. K. Li, B. Dhillon, and D. A. Mickle. 2002. Endothelin antagonism and interleukin-6 inhibition attenuate the proatherogenic effects of C-reactive protein. *Circulation*. **105**: 1890–1896.
 50. Pasceri, V., J. T. Willerson, and E. T. Yeh. 2000. Direct proinflammatory effect of C-reactive protein on human endothelial cells. *Circulation*. **102**: 2165–2168.
 51. Pasceri, V., J. S. Cheng, J. T. Willerson, E. T. Yeh, and J. Chang. 2001. Modulation of C-reactive protein-mediated monocyte chemoattractant protein-1 induction in human endothelial cells by anti-atherosclerosis drugs. *Circulation*. **103**: 2531–2534.
 52. Devaraj, S., P. R. Kumaresan, and I. Jialal. 2004. Effect of C-reactive protein on chemokine expression in human aortic endothelial cells. *J. Mol. Cell. Cardiol.* **36**: 405–410.
 53. Devaraj, S., D. Y. Xu, and I. Jialal. 2003. C-reactive protein increases plasminogen activator inhibitor-1 expression and activity in human aortic endothelial cells: implications for the metabolic syndrome and atherothrombosis. *Circulation*. **107**: 398–404.
 54. Cermak, J., N. S. Key, R. R. Bach, J. Balla, H. S. Jacob, and G. M. Vercellotti. 1993. C-reactive protein induces human peripheral blood monocytes to synthesize tissue factor. *Blood*. **82**: 513–520.

55. Tebo, J. M., and R. F. Mortensen. 1991. Internalization and degradation of receptor bound C-reactive protein by U-937 cells: induction of H₂O₂ production and tumoricidal activity. *Biochim. Biophys. Acta.* **1095**: 210–216.
56. Ballou, S. P., and G. Lozanski. 1992. Induction of inflammatory cytokine release from cultured human monocytes by C-reactive protein. *Cytokine.* **4**: 361–368.
57. Woollard, K. J., D. C. Phillips, and H. R. Griffiths. 2002. Direct modulatory effect of C-reactive protein on primary human monocyte adhesion to human endothelial cells. *Clin. Exp. Immunol.* **130**: 256–262.
58. Torzewski, M., C. Rist, R. F. Mortensen, T. P. Zwaka, M. Bienek, J. Waltenberger, W. Koenig, G. Schmitz, V. Hombach, and J. Torzewski. 2000. C-reactive protein in the arterial intima: role of C-reactive protein receptor-dependent monocyte recruitment in atherogenesis. *Arterioscler. Thromb. Vasc. Biol.* **20**: 2094–2099.
59. Williams, T. N., C. X. Zhang, B. A. Game, L. He, and Y. Huang. 2004. C-reactive protein stimulates MMP-1 expression in U937 histiocytes through Fc[gamma]RII and extracellular signal-regulated kinase pathway: an implication of CRP involvement in plaque destabilization. *Arterioscler. Thromb. Vasc. Biol.* **24**: 61–66.
60. Hattori, Y., M. Matsumura, and K. Kasai. 2003. Vascular smooth muscle cell activation by C-reactive protein. *Cardiovasc. Res.* **58**: 186–195.
61. Wang, C. H., S. H. Li, R. D. Weisel, P. W. Fedak, A. S. Dumont, P. Szmítko, R. K. Li, D. A. Mickle, and S. Verma. 2003. C-reactive protein upregulates angiotensin type 1 receptors in vascular smooth muscle. *Circulation.* **107**: 1783–1790.
62. Jialal, I., S. Devaraj, and S. K. Venugopal. 2004. C-reactive protein: risk marker or mediator in atherothrombosis? *Hypertension.* **44**: 6–11.
63. Pepys, M. B., and G. M. Hirschfield. 2003. C-reactive protein: a critical update. *J. Clin. Invest.* **111**: 1805–1812.
64. Jabs, W. J., E. Theissing, M. Nitschke, J. F. Bechtel, M. Duchrow, S. Mohamed, B. Jahrbeck, H. H. Sievers, J. Steinhoff, and C. Bartels. 2003. Local generation of C-reactive protein in diseased coronary artery venous bypass grafts and normal vascular tissue. *Circulation.* **108**: 1428–1431.
65. Zhang, Y. X., W. J. Cliff, G. I. Schoefl, and G. Higgins. 1999. Coronary C-reactive protein distribution: its relation to development of atherosclerosis. *Atherosclerosis.* **145**: 375–379.
66. Szalai, A. J. 2002. The biological functions of C-reactive protein. *Vascul. Pharmacol.* **39**: 105–107.
67. Szalai, A. J., and M. A. McCrory. 2002. Varied biologic functions of C-reactive protein: lessons learned from transgenic mice. *Immunol. Res.* **26**: 279–287.
68. Paul, A., K. W. Ko, L. Li, V. Yechoor, M. A. McCrory, A. J. Szalai, and L. Chan. 2004. C-reactive protein accelerates the progression of atherosclerosis in apolipoprotein E-deficient mice. *Circulation.* **109**: 647–655.
69. Griselli, M., J. Herbert, W. L. Hutchinson, K. M. Taylor, M. Sohail, T. Krausz, and M. B. Pepys. 1999. C-reactive protein and complement are important mediators of tissue damage in acute myocardial infarction. *J. Exp. Med.* **190**: 1733–1740.
70. Saffitz, J. E., and C. J. Schwartz. 1987. Coronary atherosclerosis and thrombosis underlying acute myocardial infarction. *Cardiol. Clin.* **5**: 21–30.
71. Nijmeijer, R., W. K. Lagrand, Y. T. Lubbers, C. A. Visser, C. J. Meijer, H. W. Niessen, and C. E. Hack. 2003. C-reactive protein activates complement in infarcted human myocardium. *Am. J. Pathol.* **163**: 269–275.
72. Coetzee, G. A., A. F. Strachan, D. R. van der Westhuyzen, H. C. Hoppe, M. S. Jeenah, and F. C. de Beer. 1986. Serum amyloid A-containing human high density lipoprotein 3. Density, size, and apolipoprotein composition. *J. Biol. Chem.* **261**: 9644–9651.
73. de Beer, M. C., T. Yuan, M. S. Kindy, B. F. Asztalos, P. S. Roheim, and F. C. de Beer. 1995. Characterization of constitutive human serum amyloid A protein (SAA4) as an apolipoprotein. *J. Lipid Res.* **36**: 526–534.
74. Whitehead, A. S., M. C. de Beer, D. M. Steel, M. Rits, J. M. Lelias, W. S. Lane, and F. C. de Beer. 1992. Identification of novel members of the serum amyloid A protein superfamily as constitutive apolipoproteins of high density lipoprotein. *J. Biol. Chem.* **267**: 3862–3867.
75. Uhlir, C. M., and A. S. Whitehead. 1999. Serum amyloid A, the major vertebrate acute-phase reactant. *Eur. J. Biochem.* **265**: 501–523.
76. Meek, R. L., N. Eriksen, and E. P. Benditt. 1989. Serum amyloid A in the mouse. Sites of uptake and mRNA expression. *Am. J. Pathol.* **135**: 411–419.
77. Meek, R. L., N. Eriksen, and E. P. Benditt. 1992. Murine serum amyloid A3 is a high density apolipoprotein and is secreted by macrophages. *Proc. Natl. Acad. Sci. USA.* **89**: 7949–7952.
78. Meek, R. L., S. Urieli-Shoval, and E. P. Benditt. 1994. Expression of apolipoprotein serum amyloid A mRNA in human atherosclerotic lesions and cultured vascular cells: implications for serum amyloid A function. *Proc. Natl. Acad. Sci. USA.* **91**: 3186–3190.
79. Yamada, T., T. Kakiyama, T. Kamishima, T. Fukuda, and T. Kawai. 1996. Both acute phase and constitutive serum amyloid A are present in atherosclerotic lesions. *Pathol. Int.* **46**: 797–800.
80. Jousilahti, P., V. Salomaa, V. Rasi, E. Vahtera, and T. Palosuo. 2001. The association of C-reactive protein, serum amyloid A and fibrinogen with prevalent coronary heart disease—baseline findings of the PAIS project. *Atherosclerosis.* **156**: 451–456.
81. Ridker, P. M., N. Rifai, M. A. Pfeffer, F. M. Sacks, L. A. Moye, S. Goldman, G. C. Flaker, and E. Braunwald. 1998. Inflammation, pravastatin, and the risk of coronary events after myocardial infarction in patients with average cholesterol levels. Cholesterol and Recurrent Events (CARE) Investigators. *Circulation.* **98**: 839–844.
82. Ridker, P. M., C. H. Hennekens, J. E. Buring, and N. Rifai. 2000. C-reactive protein and other markers of inflammation in the prediction of cardiovascular disease in women. *N. Engl. J. Med.* **342**: 836–843.
83. Erren, M., H. Reinecke, R. Junker, M. Fobker, H. Schulte, J. O. Schurek, J. Kropf, S. Kerber, G. Breithardt, G. Assmann, and P. Cullen. 1999. Systemic inflammatory parameters in patients with atherosclerosis of the coronary and peripheral arteries. *Arterioscler. Thromb. Vasc. Biol.* **19**: 2355–2363.
84. Ebeling, P., A. M. Teppo, H. A. Koistinen, J. Viikari, T. Ronnema, M. Nissen, S. Bergkulla, P. Salmela, J. Saltevo, and V. A. Koivisto. 1999. Troglitazone reduces hyperglycaemia and selectively acute-phase serum proteins in patients with type II diabetes. *Diabetologia.* **42**: 1433–1438.
85. Leinonen, E. S., A. Hiukka, E. Hurt-Camejo, O. Wiklund, S. S. Sarna, L. Mattson Hulten, J. Westerbacka, R. M. Salonen, J. T. Salonen, and M. R. Taskinen. 2004. Low-grade inflammation, endothelial activation and carotid intima-media thickness in type 2 diabetes. *J. Intern. Med.* **256**: 119–127.
86. Haffner, S., R. D'Agostino, M. Saad, D. O'Leary, P. Savage, M. Rewers, J. Selby, R. Bergman, and L. Mykkanen. 2000. Carotid artery atherosclerosis in type 2 diabetic and non-diabetic subjects with and without clinical coronary artery disease. The Insulin Resistance Atherosclerosis Study. *Am. J. Cardiol.* **85**: 1395–1400.
87. Migita, K., Y. Kawabe, M. Tominaga, T. Origuchi, T. Aoyagi, and K. Eguchi. 1998. Serum amyloid A protein induces production of matrix metalloproteinases by human synovial fibroblasts. *Lab. Invest.* **78**: 535–539.
88. Strissel, K. J., M. T. Girard, J. A. West-Mays, W. B. Rinehart, J. R. Cook, C. E. Brinckerhoff, and M. E. Fini. 1997. Role of serum amyloid A as an intermediate in the IL-1 and PMA-stimulated signaling pathways regulating expression of rabbit fibroblast collagenase. *Exp. Cell Res.* **237**: 275–287.
89. Badolato, R., J. M. Wang, W. J. Murphy, A. R. Lloyd, D. F. Michiel, L. L. Bauserman, D. J. Kelvin, and J. J. Oppenheim. 1994. Serum amyloid A is a chemoattractant: induction of migration, adhesion, and tissue infiltration of monocytes and polymorphonuclear leukocytes. *J. Exp. Med.* **180**: 203–209.
90. Xu, L., R. Badolato, W. J. Murphy, D. L. Longo, M. Anver, S. Hale, J. J. Oppenheim, and J. M. Wang. 1995. A novel biologic function of serum amyloid A. Induction of T lymphocyte migration and adhesion. *J. Immunol.* **155**: 1184–1190.
91. Artl, A., G. Marsche, S. Lestavel, W. Sattler, and E. Malle. 2000. Role of serum amyloid A during metabolism of acute-phase HDL by macrophages. *Arterioscler. Thromb. Vasc. Biol.* **20**: 763–772.
92. Liang, J., B. Schreiber, M. Salmona, G. Phillip, W. Gonnerman, F. de Beer, and J. Sipe. 1996. Amino terminal region of acute phase, but not constitutive, serum amyloid A (apoSAA) specifically binds and transports cholesterol into aortic smooth muscle and HepG2 cells. *J. Lipid Res.* **37**: 2109–2116.
93. Tam, S. P., A. Flexman, J. Hulme, and R. Kisilevsky. 2002. Promoting export of macrophage cholesterol: the physiological role of a major acute-phase protein, serum amyloid A 2.1. *J. Lipid Res.* **43**: 1410–1420.
94. Stonik, J. A., A. T. Remaley, S. J. Demosky, E. B. Neufeld, A. Bocha-

- rov, and H. B. Brewer. 2004. Serum amyloid A promotes ABCA1-dependent and ABCA1-independent lipid efflux from cells. *Biochem. Biophys. Res. Commun.* **321**: 936–941.
95. Husebekk, A., B. Skogen, and G. Husby. 1987. Characterization of amyloid proteins AA and SAA as apolipoproteins of high density lipoprotein (HDL). Displacement of SAA from the HDL-SAA complex by apo AI and apo AII. *Scand. J. Immunol.* **25**: 375–381.
96. Cabana, V. G., J. R. Lukens, K. S. Rice, T. J. Hawkins, and G. S. Getz. 1996. HDL content and composition in acute phase response in three species: triglyceride enrichment of HDL a factor in its decrease. *J. Lipid Res.* **37**: 2662–2674.
97. Ancsin, J. B., and R. Kisilevsky. 1999. The heparin/heparan sulfate-binding site on apo-serum amyloid A. Implications for the therapeutic intervention of amyloidosis. *J. Biol. Chem.* **274**: 7172–7181.
98. Ancsin, J. B., and R. Kisilevsky. 2001. Serum amyloid A peptide interactions with glycosaminoglycans. Evaluation by affinity chromatography. *Methods Mol. Biol.* **171**: 449–456.
99. Williams, K. J., and I. Tabas. 1995. The response-to-retention hypothesis of early atherogenesis. *Arterioscler. Thromb. Vasc. Biol.* **15**: 551–561.
100. Williams, K. J., and I. Tabas. 1998. The response-to-retention hypothesis of atherogenesis reinforced. *Curr. Opin. Lipidol.* **9**: 471–474.
101. Steinberg, D., S. Parthasarathy, T. E. Carew, J. C. Khoo, and J. L. Witztum. 1989. Beyond cholesterol: modifications of low density lipoprotein that increase its atherogenicity. *N. Engl. J. Med.* **320**: 916–924.
102. Lewis, K. E., E. A. Kirk, T. O. McDonald, S. Wang, T. N. Wight, K. D. O'Brien, and A. Chait. 2004. Increase in serum amyloid A evoked by dietary cholesterol is associated with increased atherosclerosis in mice. *Circulation.* **110**: 540–545.
103. Petrovic, N., C. Grove, P. E. Langton, N. L. Misso, and P. J. Thompson. 2001. A simple assay for a human serum phospholipase A2 that is associated with high-density lipoproteins. *J. Lipid Res.* **42**: 1706–1713.
104. Pruzanski, W., P. Vadas, and J. Browning. 1993. Secretory non-pancreatic group II phospholipase A2: role in physiologic and inflammatory processes. *J. Lipid Mediat.* **8**: 161–167.
105. Hurt-Camejo, E., G. Camejo, H. Peilot, K. Oorni, and P. Kovanen. 2001. Phospholipase A(2) in vascular disease. *Circ. Res.* **89**: 298–304.
106. Niessen, H. W., P. A. Krijnen, C. A. Visser, C. J. Meijer, and C. Erik Hack. 2003. Type II secretory phospholipase A2 in cardiovascular disease: a mediator in atherosclerosis and ischemic damage to cardiomyocytes? *Cardiovasc. Res.* **60**: 68–77.
107. Kugiyama, K., Y. Ota, K. Takazoe, Y. Moriyama, H. Kawano, Y. Miyao, T. Sakamoto, H. Soejima, H. Ogawa, H. Doi, S. Sugiyama, and H. Yasue. 1999. Circulating levels of secretory type II phospholipase A(2) predict coronary events in patients with coronary artery disease. *Circulation.* **100**: 1280–1284.
108. Porela, P., K. Pulkki, L. M. Voipio-Pulkki, K. Pettersson, V. Lepanen, and T. J. Nevalainen. 2000. Level of circulating phospholipase A2 in prediction of the prognosis of patients with suspected myocardial infarction. *Basic Res. Cardiol.* **95**: 413–417.
109. Austin, M. A., J. A. Breslow, C. H. Hennekens, J. E. Buring, W. C. Willett, and R. M. Krauss. 1988. Low density lipoprotein subclass pattern and risk of myocardial infarction. *J. Am. Med. Assoc.* **260**: 1917–1921.
110. Austin, M. A., M-C. King, K. M. Vranizan, and R. M. Krauss. 1990. Atherogenic lipoprotein phenotype. A proposed genetic marker for coronary heart disease risk. *Circulation.* **82**: 495–506.
111. Hurt-Camejo, E., G. Camejo, and P. Sartipy. 2000. Phospholipase A2 and small, dense low-density lipoprotein. *Curr. Opin. Lipidol.* **11**: 465–471.
112. Hurt-Camejo, E., S. Andersen, R. Standal, B. Rosengren, P. Sartipy, E. Stadberg, and B. Johansen. 1997. Localization of nonpancreatic secretory phospholipase A2 in normal and atherosclerotic arteries. Activity of the isolated enzyme on low-density lipoproteins. *Arterioscler. Thromb. Vasc. Biol.* **17**: 300–309.
113. Romano, M., E. Romano, S. Bjorkerud, and E. Hurt-Camejo. 1998. Ultrastructural localization of secretory type II phospholipase A2 in atherosclerotic and nonatherosclerotic regions of human arteries. *Arterioscler. Thromb. Vasc. Biol.* **18**: 519–525.
114. Sartipy, P., G. Bondjers, and E. Hurt-Camejo. 1998. Phospholipase A2 type II binds to extracellular matrix biglycan: modulation of its activity on LDL by colocalization in glycosaminoglycan matrixes. *Arterioscler. Thromb. Vasc. Biol.* **18**: 1934–1941.
115. Hakala, J. K., K. Oorni, M. Ala-Korpela, and P. T. Kovanen. 1999. Lipolytic modification of LDL by phospholipase A2 induces particle aggregation in the absence and fusion in the presence of heparin. *Arterioscler. Thromb. Vasc. Biol.* **19**: 1276–1283.
116. Sartipy, P., G. Camejo, L. Svensson, and E. Hurt-Camejo. 1999. Phospholipase A(2) modification of low density lipoproteins forms small high density particles with increased affinity for proteoglycans and glycosaminoglycans. *J. Biol. Chem.* **274**: 25913–25920.
117. Heinecke, J. W., A. G. Suits, M. Aviram, and A. Chait. 1991. Phagocytosis of lipase-aggregated low density lipoprotein promotes macrophage foam cell formation. *Arterioscler. Thromb.* **11**: 1643–1651.
118. Suits, A. G., A. Chait, M. Aviram, and J. W. Heinecke. 1989. Phagocytosis of aggregated lipoprotein by macrophages: low density lipoprotein receptor-dependent foam cell formation. *Proc. Natl. Acad. Sci. USA.* **86**: 2713–2717.
119. Khoo, J. C., E. Miller, P. McLoughlin, and D. Steinberg. 1988. Enhanced macrophage uptake of low density lipoprotein after self-aggregation. *Arteriosclerosis.* **8**: 348–358.
120. Quinn, M. T., S. Parthasarathy, and D. Steinberg. 1988. Lysophosphatidylcholine: a chemotactic factor for human monocytes and its potential role in atherogenesis. *Proc. Natl. Acad. Sci. USA.* **85**: 2805–2809.
121. Chang, M. Y., C. Tsoi, T. N. Wight, and A. Chait. 2003. Lysophosphatidylcholine regulates synthesis of biglycan and the proteoglycan form of macrophage colony stimulating factor. *Arterioscler. Thromb. Vasc. Biol.* **23**: 809–815.
122. Kikuchi, A., H. Tomoyasu, I. Kido, K. Takahashi, A. Tanaka, I. Nonaka, N. Iwakami, and I. Kamo. 2000. Haemopoietic biglycan produced by brain cells stimulates growth of microglial cells. *J. Neuroimmunol.* **106**: 78–86.
123. Sakai, M., A. Miyazaki, H. Hakamata, T. Sasaki, S. Yui, M. Yamazaki, M. Shichiri, and S. Horiuchi. 1994. Lysophosphatidylcholine plays an essential role in the mitogenic effect of oxidized low density lipoprotein on murine macrophages. *J. Biol. Chem.* **269**: 31430–31435.
124. Sakai, M., A. Miyazaki, H. Hakamata, S. Kobori, M. Shichiri, and S. Horiuchi. 1996. Endocytic uptake of lysophosphatidylcholine mediated by macrophage scavenger receptor plays a major role in oxidized low density lipoprotein-induced macrophage growth. *J. Atheroscler. Thromb.* **2**: 81–86.
125. Kume, N., M. I. Cybulsky, and M. A. J. Gimbrone. 1992. Lysophosphatidylcholine, a component of atherogenic lipoproteins, induces mononuclear leukocyte adhesion molecules in cultured human and rabbit arterial endothelial cells. *J. Clin. Invest.* **90**: 1138–1144.
126. Yokote, K., N. Morisaki, M. Zenibayashi, S. Ueda, T. Kanzaki, Y. Saito, and S. Yoshida. 1993. The phospholipase-A2 reaction leads to increased monocyte adhesion of endothelial cells via the expression of adhesion molecules. *Eur. J. Biochem.* **217**: 723–729.
127. Nakano, T., E. W. Raines, J. A. Abraham, M. Klagsbrun, and R. Ross. 1994. Lysophosphatidylcholine upregulates the level of heparin-binding epidermal growth factor-like growth factor mRNA in human monocytes. *Proc. Natl. Acad. Sci. USA.* **91**: 1069–1073.
128. Kume, N., and M. A. J. Gimbrone. 1994. Lysophosphatidylcholine transcriptionally induces growth factor gene expression in cultured human endothelial cells. *J. Clin. Invest.* **93**: 907–911.
129. Nishi, E., N. Kume, H. Ochi, H. Moriwaki, Y. Wakatsuki, S. Higashiyama, N. Taniguchi, and T. Kita. 1997. Lysophosphatidylcholine increases expression of heparin-binding epidermal growth factor-like growth factor in human T lymphocytes. *Circ. Res.* **80**: 638–644.
130. Simon, M. F., H. Chap, and L. Douste-Blazy. 1984. Platelet aggregating activity of lysophosphatidic acids is not related to their calcium ionophore properties. *FEBS Lett.* **166**: 115–119.
131. Haseruck, N., W. Erl, D. Pandey, G. Tigyi, P. Ohlmann, C. Ravanat, C. Gachet, and W. Siess. 2004. The plaque lipid lysophosphatidic acid stimulates platelet activation and platelet-monocyte aggregate formation in whole blood: involvement of P2Y1 and P2Y12 receptors. *Blood.* **103**: 2585–2592.
132. MacPhee, M., K. P. Chepenik, R. A. Liddell, K. K. Nelson, L. D. Sircusa, and A. M. Buchberg. 1995. The secretory phospholipase A2 gene is a candidate for the Mom1 locus, a major modifier of Apc-Min-induced intestinal neoplasia. *Cell.* **81**: 957–966.
133. Webb, N. R., M. A. Bostrom, S. J. Szilvassy, D. R. van der Westhuyzen, A. Daugherty, and F. C. de Beer. 2003. Macrophage-expressed group IIA secretory phospholipase A2 increases atherosclerotic lesion formation in LDL receptor-deficient mice. *Arterioscler. Thromb. Vasc. Biol.* **23**: 263–268.

134. Ivandic, B., L. W. Castellani, X. P. Wang, J. H. Qiao, M. Mehrabian, M. Navab, A. M. Fogelman, D. S. Grass, M. E. Swanson, M. C. de Beer, F. de Beer, and A. J. Lusis. 1999. Role of group II secretory phospholipase A2 in atherosclerosis. I. Increased atherogenesis and altered lipoproteins in transgenic mice expressing group IIa phospholipase A2. *Arterioscler. Thromb. Vasc. Biol.* **19**: 1284–1290.
135. Tietge, U. J., C. Maugeais, S. Lund-Katz, D. Grass, F. C. de Beer, and D. J. Rader. 2002. Human secretory phospholipase A2 mediates decreased plasma levels of HDL cholesterol and apoA-I in response to inflammation in human apoA-I transgenic mice. *Arterioscler. Thromb. Vasc. Biol.* **22**: 1213–1218.
136. Burton, C. A., S. Patel, S. Mundt, H. Hassing, D. Zhang, A. Hermanowski-Vosatka, S. D. Wright, Y. S. Chao, P. A. Detmers, and C. P. Sparrow. 2002. Deficiency in sPLA(2) does not affect HDL levels or atherosclerosis in mice. *Biochem. Biophys. Res. Commun.* **294**: 88–94.
137. Murakami, M., and I. Kudo. 2003. New phospholipase A(2) isozymes with a potential role in atherosclerosis. *Curr. Opin. Lipidol.* **14**: 431–436.
138. Gesquiere, L., W. Cho, and P. V. Subbiah. 2002. Role of group IIa and group V secretory phospholipases A(2) in the metabolism of lipoproteins. Substrate specificities of the enzymes and the regulation of their activities by sphingomyelin. *Biochemistry.* **41**: 4911–4920.
139. Suckling, K. E., and C. H. Macphee. 2002. Lipoprotein-associated phospholipase A2: a target directed at the atherosclerotic plaque. *Expert Opin. Ther. Targets.* **6**: 309–314.
140. Caslake, M. J., and C. J. Packard. 2003. Lipoprotein-associated phospholipase A2 (platelet-activating factor acetylhydrolase) and cardiovascular disease. *Curr. Opin. Lipidol.* **14**: 347–352.
141. Chen, C. H. 2004. Platelet-activating factor acetylhydrolase: is it good or bad for you? *Curr. Opin. Lipidol.* **15**: 337–341.
142. Memon, R. A., J. Fuller, A. H. Moser, K. R. Feingold, and C. Grunfeld. 1999. In vivo regulation of plasma platelet-activating factor acetylhydrolase during the acute phase response. *Am. J. Physiol.* **277**: R94–R103.
143. Van Lenten, B. J., S. Y. Hama, F. C. de Beer, D. M. Stafforini, T. M. McIntyre, S. M. Prescott, B. N. La Du, A. M. Fogelman, and M. Navab. 1995. Anti-inflammatory HDL becomes pro-inflammatory during the acute phase response. Loss of protective effect of HDL against LDL oxidation in aortic wall cell cocultures. *J. Clin. Invest.* **96**: 2758–2767.
144. Boisfer, E., D. Stengel, D. Pastier, P. M. Laplaud, N. Dousset, E. Ninio, and A. D. Kalopissis. 2002. Antioxidant properties of HDL in transgenic mice overexpressing human apolipoprotein A-II. *J. Lipid Res.* **43**: 732–741.
145. Packard, C. J., D. S. O'Reilly, M. J. Caslake, A. D. McMahon, I. Ford, J. Cooney, C. H. Macphee, K. E. Suckling, M. Krishna, F. E. Wilkinson, A. Rumley, and G. D. Lowe. 2000. Lipoprotein-associated phospholipase A2 as an independent predictor of coronary heart disease. West of Scotland Coronary Prevention Study Group. *N. Engl. J. Med.* **343**: 1148–1155.
146. Koenig, W., N. Khuseynova, H. Lowel, G. Trischler, and C. Meisinger. 2004. Lipoprotein-associated phospholipase A2 adds to risk prediction of incident coronary events by C-reactive protein in apparently healthy middle-aged men from the general population: results from the 14-year follow-up of a large cohort from southern Germany. *Circulation.* **110**: 1903–1908.
147. Ballantyne, C. M., R. C. Hoogeveen, H. Bang, J. Coresh, A. R. Folsom, G. Heiss, and A. R. Sharrett. 2004. Lipoprotein-associated phospholipase A2, high-sensitivity C-reactive protein, and risk for incident coronary heart disease in middle-aged men and women in the Atherosclerosis Risk in Communities (ARIC) Study. *Circulation.* **109**: 837–842.
148. Llodra, J., V. Angeli, J. Liu, E. Trogan, E. A. Fisher, and G. J. Randolph. 2004. Emigration of monocyte-derived cells from atherosclerotic lesions characterizes regressive, but not progressive, plaques. *Proc. Natl. Acad. Sci. USA.* **101**: 11779–11784.
149. Angeli, V., J. Llodra, J. X. Rong, K. Satoh, S. Ishii, T. Shimizu, E. A. Fisher, and G. J. Randolph. 2004. Dyslipidemia associated with atherosclerotic disease systemically alters dendritic cell mobilization. *Immunity.* **21**: 561–574.
150. Forte, T. M., G. Subbanagounder, J. A. Berliner, P. J. Blanche, A. O. Clermont, Z. Jia, M. N. Oda, R. M. Krauss, and J. K. Bielicki. 2002. Altered activities of anti-atherogenic enzymes LCAT, paraoxonase, and platelet-activating factor acetylhydrolase in atherosclerosis-susceptible mice. *J. Lipid Res.* **43**: 477–485.
151. Hase, M., M. Tanaka, M. Yokota, and Y. Yamada. 2002. Reduction in the extent of atherosclerosis in apolipoprotein E-deficient mice induced by electroporation-mediated transfer of the human plasma platelet-activating factor acetylhydrolase gene into skeletal muscle. *Prostaglandins Other Lipid Mediat.* **70**: 107–118.
152. Costa, L. G., B. E. McDonald, S. D. Murphy, G. S. Omenn, R. J. Richter, A. G. Motulsky, and C. E. Furlong. 1990. Serum paraoxonase and its influence on paraoxon and chlorpyrifos-oxon toxicity in rats. *Toxicol. Appl. Pharmacol.* **103**: 66–76.
153. La Du, B. N. 1992. Human serum paraoxonase/arylesterase. In *Pharmacogenetics of Drug Metabolism*. W. Kalow, editor. Pergamon Press, Inc., New York. 51–91.
154. Bergmeier, C., R. Siekmeier, and W. Gross. 2004. Distribution spectrum of paraoxonase activity in HDL fractions. *Clin. Chem.* **50**: 2309–2315.
155. Ng, C. J., D. J. Wadleigh, A. Gangopadhyay, S. Hama, V. R. Grijalva, M. Navab, A. M. Fogelman, and S. T. Reddy. 2001. Paraoxonase-2 is a ubiquitously expressed protein with antioxidant properties and is capable of preventing cell-mediated oxidative modification of low density lipoprotein. *J. Biol. Chem.* **276**: 44444–44449.
156. Draganov, D. I., P. L. Stetson, C. E. Watson, S. S. Billecke, and B. N. La Du. 2000. Rabbit serum paraoxonase 3 (PON3) is a high density lipoprotein-associated lactonase and protects low density lipoprotein against oxidation. *J. Biol. Chem.* **275**: 33435–33442.
157. Reddy, S. T., D. J. Wadleigh, V. Grijalva, C. Ng, S. Hama, A. Gangopadhyay, D. M. Shih, A. J. Lusis, M. Navab, and A. M. Fogelman. 2001. Human paraoxonase-3 is an HDL-associated enzyme with biological activity similar to paraoxonase-1 protein but is not regulated by oxidized lipids. *Arterioscler. Thromb. Vasc. Biol.* **21**: 542–547.
158. Cabana, V. G., C. A. Reardon, N. Feng, S. Neath, J. Lukens, and G. S. Getz. 2003. Serum paraoxonase: effect of the apolipoprotein composition of HDL and the acute phase response. *J. Lipid Res.* **44**: 780–792.
159. Feingold, K. R., R. A. Memon, A. H. Moser, and C. Grunfeld. 1998. Paraoxonase activity in the serum and hepatic mRNA levels decrease during the acute phase response. *Atherosclerosis.* **139**: 307–315.
160. Kumon, Y., Y. Nakauchi, K. Kidawara, M. Fukushima, S. Kobayashi, Y. Ikeda, T. Suehiro, K. Hashimoto, and J. D. Sipe. 1998. A longitudinal analysis of alteration in lecithin-cholesterol acyltransferase and paraoxonase activities following laparoscopic cholecystectomy relative to other parameters of HDL function and the acute phase response. *Scand. J. Immunol.* **48**: 419–424.
161. Navab, M., S. Hama-Levy, B. J. Van Lenten, G. C. Fonarow, C. J. Cardinez, L. W. Castellani, M. L. Brennan, A. J. Lusis, A. M. Fogelman, and B. N. La Du. 1997. Mildly oxidized LDL induces an increased apolipoprotein J/paraoxonase ratio. *J. Clin. Invest.* **99**: 2005–2019.
162. Shih, D. M., L. Gu, S. Hama, Y. R. Xia, M. Navab, A. M. Fogelman, and A. J. Lusis. 1996. Genetic-dietary regulation of serum paraoxonase expression and its role in atherogenesis in a mouse model. *J. Clin. Invest.* **97**: 1630–1639.
163. La Du, B. N. 1988. Invited editorial. The human serum paraoxonase/arylesterase polymorphism. *Am. J. Hum. Genet.* **43**: 227–229.
164. Li, H. L., D. P. Liu, and C. C. Liang. 2003. Paraoxonase gene polymorphisms, oxidative stress, and diseases. *J. Mol. Med.* **81**: 766–779.
165. Mackness, M., P. Durrington, and B. Mackness. 2004. Paraoxonase 1 activity, concentration and genotype in cardiovascular disease. *Curr. Opin. Lipidol.* **15**: 399–404.
166. Barter, P. J., S. Nicholls, K. A. Rye, G. M. Anantharamaiah, M. Navab, and A. M. Fogelman. 2004. Antiinflammatory properties of HDL. *Circ. Res.* **95**: 764–772.
167. Mackness, M. I., S. Arrol, C. Abbott, and P. N. Durrington. 1993. Protection of low-density lipoprotein against oxidative modification by high-density lipoprotein associated paraoxonase. *Arteriosclerosis.* **104**: 129–135.
168. Mackness, M. I., S. Arrol, and P. N. Durrington. 1991. Paraoxonase prevents accumulation of lipoperoxides in low-density lipoproteins. *FEBS Lett.* **286**: 152–154.
169. Navab, M., S. Y. Hama, G. P. Hough, C. C. Hedrick, R. Sorenson, B. N. La Du, J. A. Kobashigawa, G. C. Fonarow, J. A. Berliner, H. Laks, and A. M. Fogelman. 1998. High density associated enzymes: their role in vascular biology. *Curr. Opin. Lipidol.* **9**: 449–456.
170. Navab, M., G. M. Anantharamaiah, S. T. Reddy, B. J. Van Lenten, B. J. Ansell, G. C. Fonarow, K. Vahabzadeh, S. Hama, G. Hough, N. Kamranpour, J. A. Berliner, A. J. Lusis, and A. M. Fogelman. 2004.

- The oxidation hypothesis of atherogenesis: the role of oxidized phospholipids and HDL. *J. Lipid Res.* **45**: 993–1007.
171. Witztum, J. L., and D. Steinberg. 1991. Role of oxidized low density lipoprotein in atherogenesis. *J. Clin. Invest.* **88**: 1785–1792.
172. Parthasarathy, S., J. Barnett, and L. G. Fong. 1990. High-density lipoprotein inhibits the oxidative modification of low-density lipoprotein. *Biochim. Biophys. Acta.* **1044**: 275–283.
173. Shih, D. M., Y. R. Xia, X. P. Wang, E. Miller, L. W. Castellani, G. Subbanagounder, H. Cheroutre, K. F. Faull, J. A. Berliner, J. L. Witztum, and A. J. Lusis. 2000. Combined serum paraoxonase knockout/apolipoprotein E knockout mice exhibit increased lipoprotein oxidation and atherosclerosis. *J. Biol. Chem.* **275**: 17527–17535.
174. Rozenberg, O., M. Rosenblat, R. Coleman, D. M. Shih, and M. Aviram. 2003. Paraonase (PON1) deficiency is associated with increased macrophage oxidative stress: studies in PON1-knockout mice. *Free Radic. Biol. Med.* **34**: 774–784.
175. Tward, A., Y. R. Xia, X. P. Wang, Y. S. Shi, C. Park, L. W. Castellani, A. J. Lusis, and D. M. Shih. 2002. Decreased atherosclerotic lesion formation in human serum paraoxonase transgenic mice. *Circulation.* **106**: 484–490.
176. Navab, M., J. A. Berliner, A. D. Watson, S. Y. Hama, M. C. Territo, A. J. Lusis, D. M. Shih, B. J. Van Lenten, J. S. Frank, L. L. Demer, P. A. Edwards, and A. M. Fogelman. 1996. The yin and yang of oxidation in the development of the fatty streak. A review based on the 1994 George Lyman Duff Memorial Lecture. *Arterioscler. Thromb. Vasc. Biol.* **16**: 831–842.
177. Marathe, G. K., G. A. Zimmerman, and T. M. McIntyre. 2003. Platelet-activating factor acetylhydrolase, and not paraoxonase-1, is the oxidized phospholipid hydrolase of high density lipoprotein particles. *J. Biol. Chem.* **278**: 3937–3947.
178. Teiber, J. F., D. I. Draganov, and B. N. La Du. 2004. Purified human serum PON1 does not protect LDL against oxidation in the in vitro assays initiated with copper or AAPH. *J. Lipid Res.* **45**: 2260–2268.
179. Jordan-Starck, T. C., S. D. Lund, D. P. Witte, B. J. Aronow, C. A. Ley, W. D. Stuart, D. K. Swertfeger, L. R. Clayton, S. F. Sells, B. Paigen, and J. A. Harmony. 1994. Mouse apolipoprotein J: characterization of a gene implicated in atherosclerosis. *J. Lipid Res.* **35**: 194–210.
180. Ishikawa, Y., Y. Akasaka, T. Ishii, K. Komiyama, S. Masuda, N. Asuwa, N. H. Choi-Miura, and M. Tomita. 1998. Distribution and synthesis of apolipoprotein J in the atherosclerotic aorta. *Arterioscler. Thromb. Vasc. Biol.* **18**: 665–672.
181. Olin, K. L., S. Potter-Perigo, P. H. Barrett, T. N. Wight, and A. Chait. 2001. Biglycan, a vascular proteoglycan, binds differently to HDL(2) and HDL(3): role of apoE. *Arterioscler. Thromb. Vasc. Biol.* **21**: 129–135.
182. Olin-Lewis, K., J. Benton, J. Rutledge, D. Baskin, T. Wight, and A. Chait. 2002. Apolipoprotein E mediates the retention of high-density lipoproteins by mouse carotid arteries and cultured arterial smooth muscle cell extracellular matrices. *Circ. Res.* **90**: 1333–1339.
183. Witte, D. P., B. J. Aronow, M. L. Stauderman, W. D. Stuart, M. A. Clay, R. A. Gruppo, S. H. Jenkins, and J. A. Harmony. 1993. Platelet activation releases megakaryocyte-synthesized apolipoprotein J, a highly abundant protein in atheromatous lesions. *Am. J. Pathol.* **143**: 763–773.
184. Rosenber, M. E., and J. Silkensen. 1995. Clusterin: physiologic and pathophysiological considerations. *Int. J. Biochem. Cell Biol.* **27**: 633–645.
185. Trougakos, I. P., M. Poulakou, M. Stathatos, A. Chalikia, A. Melidonis, and E. S. Gonos. 2002. Serum levels of the senescence biomarker clusterin/apolipoprotein J increase significantly in diabetes type II and during development of coronary heart disease or at myocardial infarction. *Exp. Gerontol.* **37**: 1175–1187.
186. Mackness, B., G. K. Davies, W. Turkie, E. Lee, D. H. Roberts, E. Hill, C. Roberts, P. N. Durrington, and M. I. Mackness. 2001. Paraonase status in coronary heart disease: are activity and concentration more important than genotype? *Arterioscler. Thromb. Vasc. Biol.* **21**: 1451–1457.
187. Mackness, B., P. Durrington, P. McElduff, J. Yarnell, N. Azam, M. Watt, and M. Mackness. 2003. Low paraonase activity predicts coronary events in the Caerphilly Prospective Study. *Circulation.* **107**: 2775–2779.
188. Cabana, V. G., J. N. Siegel, and S. M. Sabesin. 1989. Effects of the acute phase response on the concentration and density distribution of plasma lipids and apolipoproteins. *J. Lipid Res.* **30**: 39–49.
189. Pruzanski, W., E. Stefanski, F. C. de Beer, M. C. de Beer, A. Ravandi, and A. Kuksis. 2000. Comparative analysis of lipid composition of normal and acute-phase high density lipoproteins. *J. Lipid Res.* **41**: 1035–1047.
190. König, V., U. Hopf, B. Moller, H. Lobeck, G. Assmann, M. Freudenberg, and C. Galanos. 1988. The significance of high-density lipoproteins (HDL) in the clearance of intravenously administered bacterial lipopolysaccharides (LPS) in mice. *Hepatogastroenterology.* **35**: 111–115.
191. Wu, A., C. J. Hinds, and C. Thiemermann. 2004. High-density lipoproteins in sepsis and septic shock: metabolism, actions, and therapeutic applications. *Shock.* **21**: 210–221.
192. Miller, G. J. 1980. HDL and atherosclerosis. *Annu. Rev. Med.* **31**: 97–108.
193. Gordon, D. J., J. Knoke, J. L. Probstfield, R. Superko, and H. A. Tyroler. 1986. High-density lipoprotein cholesterol and coronary artery disease in hypercholesterolemic men. The Lipid Research Clinics Coronary Primary Prevention Trial. *Circulation.* **73**: 1217–1225.
194. Gordon, T., W. P. Castelli, M. C. Hjortland, W. B. Kannel, and T. R. Dawber. 1977. High density lipoproteins as a protective factor against coronary heart disease. The Framingham Study. *Am. J. Med.* **62**: 707–714.
195. Grundy, S. M., H. B. Brewer, Jr., J. I. Cleeman, S. C. Smith, Jr., and C. Lenfant. 2004. Definition of metabolic syndrome: report of the National Heart, Lung, and Blood Institute/American Heart Association conference on scientific issues related to definition. *Arterioscler. Thromb. Vasc. Biol.* **24**: e13–e18.
196. Bergt, C., S. Pennathur, X. Fu, J. Byun, K. O'Brien, T. O. McDonald, P. Singh, G. M. Anantharamaiah, A. Chait, J. Brunzell, R. L. Geary, J. F. Oram, and J. W. Heinecke. 2004. The myeloperoxidase product hypochlorous acid oxidizes HDL in the human artery wall and impairs ABCA1-dependent cholesterol transport. *Proc. Natl. Acad. Sci. USA.* **101**: 13032–13037.
197. Pennathur, S., C. Bergt, B. Shao, J. Byun, S. Y. Kassim, P. Singh, P. S. Green, T. O. McDonald, J. Brunzell, A. Chait, J. F. Oram, K. O'Brien, R. L. Geary, and J. W. Heinecke. 2004. Human atherosclerotic intima and blood of patients with established coronary artery disease contain high density lipoprotein damaged by reactive nitrogen species. *J. Biol. Chem.* **279**: 42977–42983.
198. Zheng, L., B. Nukuna, M. L. Brennan, M. Sun, M. Goormastic, M. Settle, D. Schmitt, X. Fu, L. Thomson, P. L. Fox, H. Ischiropoulos, J. D. Smith, M. Kinter, and S. L. Hazen. 2004. Apolipoprotein A-I is a selective target for myeloperoxidase-catalyzed oxidation and functional impairment in subjects with cardiovascular disease. *J. Clin. Invest.* **114**: 529–541.
199. Zheng, L., M. Settle, G. Brubaker, D. Schmitt, S. L. Hazen, J. D. Smith, and M. Kinter. 2005. Localization of nitration and chlorination sites on apolipoprotein A-I catalyzed by myeloperoxidase in human atheroma and associated oxidative impairment in ABCA1-dependent cholesterol efflux from macrophages. *J. Biol. Chem.* **280**: 38–47.
200. Bergt, C., X. Fu, N. P. Huq, J. Kao, and J. W. Heinecke. 2004. Lysine residues direct the chlorination of tyrosines in YXXK motifs of apolipoprotein A-I when hypochlorous acid oxidizes high density lipoprotein. *J. Biol. Chem.* **279**: 7856–7866.
201. Brennan, M. L., M. M. Anderson, D. M. Shih, X. D. Qu, X. Wang, A. C. Mehta, L. L. Lim, W. Shi, S. L. Hazen, J. S. Jacob, J. R. Crowley, J. W. Heinecke, and A. J. Lusis. 2001. Increased atherosclerosis in myeloperoxidase-deficient mice. *J. Clin. Invest.* **107**: 419–430.
202. Gaut, J. P., J. Byun, H. D. Tran, W. M. Lauber, J. A. Carroll, R. S. Hotchkiss, A. Belaouaj, and J. W. Heinecke. 2002. Myeloperoxidase produces nitrating oxidants in vivo. *J. Clin. Invest.* **109**: 1311–1319.
203. Lawn, R. M., D. P. Wade, M. R. Garvin, X. Wang, K. Schwartz, J. G. Porter, J. J. Seilhamer, A. M. Vaughan, and J. F. Oram. 1999. The Tangier disease gene product ABC1 controls the cellular apolipoprotein-mediated lipid removal pathway. *J. Clin. Invest.* **104**: R25–R31.
204. Oram, J., and R. Lawn. 2001. ABCA1: the gatekeeper for eliminating excess tissue cholesterol. *J. Lipid Res.* **42**: 1173–1179.
205. Brewer, H. B., Jr., and S. Santamarina-Fojo. 2003. New insights into the role of the adenosine triphosphate-binding cassette transporters in high-density lipoprotein metabolism and reverse cholesterol transport. *Am. J. Cardiol.* **91**: 3E–11E.
206. Fluiter, K., H. Vietsch, E. A. Biessen, G. M. Kostner, T. J. van Ber-

- kel, and W. Sattler. 1996. Increased selective uptake in vivo and in vitro of oxidized cholesteryl esters from high-density lipoprotein by rat liver parenchymal cells. *Biochem. J.* **319**: 471–476.
207. Barter, P. J., P. W. Baker, and K. A. Rye. 2002. Effect of high-density lipoproteins on the expression of adhesion molecules in endothelial cells. *Curr. Opin. Lipidol.* **13**: 285–288.
208. Cockerill, G. W., K. A. Rye, J. R. Gamble, M. A. Vadas, and P. J. Barter. 1995. High-density lipoproteins inhibit cytokine-induced expression of endothelial cell adhesion molecules. *Arterioscler. Thromb. Vasc. Biol.* **15**: 1987–1994.
209. Wadham, C., N. Albanese, J. Roberts, L. Wang, C. J. Bagley, J. R. Gamble, K. A. Rye, P. J. Barter, M. A. Vadas, and P. Xia. 2004. High-density lipoproteins neutralize C-reactive protein proinflammatory activity. *Circulation.* **109**: 2116–2122.
210. Shao, B., C. Bergt, X. Fu, P. Green, J. C. Voss, M. N. Oda, J. F. Oram, and J. W. Heinecke. 2004. Tyrosine 192 in apolipoprotein A-I is the major site of nitration and chlorination by myeloperoxidase, but only chlorination markedly impairs ABCA1-dependent cholesterol transport. *J. Biol. Chem.* In press.
211. Weisberg, S. P., D. McCann, M. Desai, M. Rosenbaum, R. L. Leibel, and A. W. Ferrante, Jr. 2003. Obesity is associated with macrophage accumulation in adipose tissue. *J. Clin. Invest.* **112**: 1796–1808.
212. Xu, H., G. T. Barnes, Q. Yang, G. Tan, D. Yang, C. J. Chou, J. Sole, A. Nichols, J. S. Ross, L. A. Tartaglia, and H. Chen. 2003. Chronic inflammation in fat plays a crucial role in the development of obesity-related insulin resistance. *J. Clin. Invest.* **112**: 1821–1830.
213. You, T., R. Yang, M. F. Lyles, D. Gong, and B. J. Nicklas. 2004. Abdominal adipose tissue cytokine gene expression: relationship to obesity and metabolic risk factors. *Am. J. Physiol. Endocrinol. Metab.* In press.
214. Wisse, B. E. 2004. The inflammatory syndrome: the role of adipose tissue cytokines in metabolic disorders linked to obesity. *J. Am. Soc. Nephrol.* **15**: 2792–2800.
215. Lin, Y., M. W. Rajala, J. P. Berger, D. E. Moller, N. Barzilay, and P. E. Scherer. 2001. Hyperglycemia-induced production of acute phase reactants in adipose tissue. *J. Biol. Chem.* **276**: 42077–42083.
216. Grayston, J. T. 2000. Background and current knowledge of *Chlamydia pneumoniae* and atherosclerosis. *J. Infect. Dis.* **181**(Suppl. 3): 402–410.
217. Chiu, B., E. Viira, W. Tucker, and I. W. Fong. 1997. *Chlamydia pneumoniae*, cytomegalovirus, and herpes simplex virus in atherosclerosis of the carotid artery. *Circulation.* **96**: 2144–2148.
218. Mayr, M., S. Kiechl, J. Willeit, G. Wick, and Q. Xu. 2000. Infections, immunity, and atherosclerosis: associations of antibodies to *Chlamydia pneumoniae*, *Helicobacter pylori*, and cytomegalovirus with immune reactions to heat-shock protein 60 and carotid or femoral atherosclerosis. *Circulation.* **102**: 833–839.
219. Benditt, E. P., T. Barrett, and J. K. McDougall. 1983. Viruses in the etiology of atherosclerosis. *Proc. Natl. Acad. Sci. USA.* **80**: 6386–6389.
220. Khovidhunkit, W., M. S. Kim, R. A. Memon, J. K. Shigenaga, A. H. Moser, K. R. Feingold, and C. Grunfeld. 2004. Effects of infection and inflammation on lipid and lipoprotein metabolism: mechanisms and consequences to the host. *J. Lipid Res.* **45**: 1169–1196.
221. Yoo, J. Y., and S. Desiderio. 2003. Innate and acquired immunity intersect in a global view of the acute-phase response. *Proc. Natl. Acad. Sci. USA.* **100**: 1157–1162.
222. Batista, M. C., F. K. Welty, M. R. Diffenderfer, M. J. Sarnak, E. J. Schaefer, S. Lamon-Fava, B. F. Asztalos, G. G. Dolnikowski, M. E. Brousseau, and J. B. Marsh. 2004. Apolipoprotein A-I, B-100, and B-48 metabolism in subjects with chronic kidney disease, obesity, and the metabolic syndrome. *Metabolism.* **53**: 1255–1261.
223. Park, S. H., W. Y. Lee, Y. S. Lee, E. J. Rhee, and S. W. Kim. 2004. The relative effects of obesity and insulin resistance on cardiovascular risk factors in nondiabetic and normotensive men. *Korean J. Intern. Med.* **19**: 75–80.
224. Couillard, C., B. Lamarche, A. Tchernof, D. Prud'homme, A. Tremblay, C. Bouchard, S. Moorjani, A. Nadeau, P. J. Lupien, and J. P. Despres. 1996. Plasma high-density lipoprotein cholesterol but not apolipoprotein A-I is a good correlate of the visceral obesity-insulin resistance dyslipidemic syndrome. *Metabolism.* **45**: 882–888.
225. Morishima, A., N. Ohkubo, N. Maeda, T. Miki, and N. Mitsuda. 2003. NFκB regulates plasma apolipoprotein A-I and high density lipoprotein cholesterol through inhibition of peroxisome proliferator-activated receptor alpha. *J. Biol. Chem.* **278**: 38188–38193.
226. Han, C., and A. Chait. 2004. Apolipoprotein A-I and serum amyloid A are reciprocally regulated by inflammation: potential role in formation of proatherogenic HDL. *Circulation.* **110**: III-36.